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# Dipeptidyl peptidase 4 inhibitor improves insulin resistance in Japanese patients with type 2 diabetes: a single-arm study, a brief report

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# **Abstract**

**Background:** Dipeptidyl peptidase 4 inhibitor (DPP4i) is an effective medicine for type 2 diabetes mellitus (T2DM). Some articles reported DPP4i improves insulin secretion and insulin resistance. However, these effects are not well established by glucose clamp test and test meal in Japanese. We investigated the effect of DPP4i on insulin resistance and insulin secretion by using the glucose clamp test and meal tolerance test (MTT).

**Methods:** We performed a MTT, and the hyperinsulinemic-euglycemic clamp in 8 Japanese patients with T2DM. This study was a single-arm study. We measured fasting and postprandial glucose, insulin, incretins, and glucagon levels. We also measured serum adiponectin levels.

**Results:** HbA1c was significantly decreased after 3 months. The fasting and postprandial glucose levels were significantly decreased. Fasting and postprandial insulin levels were not changed. The insulin resistance derived from the glucose clamp test was significantly improved. HOMA-IR was not significantly changed. GLP-1 and GIP were significantly increased but glucagon did not change. Adiponectin was not significantly changed.

**Conclusions:** Although the number of patients was very small, these results suggested that DPP4i treatment might improve insulin resistance without changing insulin secretion.

**Keywords:** Dipeptidyl peptidase 4 inhibitor, Insulin Resistance, Type 2 Diabetes, Glucose Clamp

# Introduction

The major pathophysiology of type 2 diabetes mellitus (T2DM) is increased insulin resistance and decreased insulin secretion [1]. Dipeptidyl dipeptidase inhibitor 4 inhibitors (DDP4i) are an anti-diabetic medicine, which increases incretin hormones, glucagon-like peptide-1 (GLP-1), glucose-dependent insulinotropic polypeptide (GIP), induce glucose-lowering effect [2]. DPP-4

inhibitors were firstly reported that improved glucose levels, increased insulin secretion, and improved betacell function in rodents [3]. However, a human study reported that DPP-4 inhibitor decreased fasting and postprandial plasma glucose, and HbA1c without changing fasting and postprandial insulin in T2DM patients [4]. Another study reported that sitagliptin increased post-load insulin and decreased post-load glucagon at 2 h in the oral glucose tolerance test (OGTT) compared with placebo [5]. We also previously reported sitagliptin improved postprandial glucose and early insulin response, however, the insulin secretion enhancing effects were very mild [6]. Previous another study

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reported that DPP4i improve insulin resistance evaluated by glucose clamp test in American, but insulin secretion did not change [7]. However, there are few reports about the effect of DPP4i on insulin resistance evaluated by the glucose clamp test in Asian and Japanese populations. East Asian and American show different pathophysiology of T2DM, East Asian shows insulin resistance with mild obesity and low insulin secretion ability [8]. Furthermore, there is a report that DPP4i increases serum adiponectin levels [9]. Adiponectin is important for insulin resistance in the Japanese population [10]. East Asians (Japanese, China, Korean, Taiwanese) showed a significantly greater effect of DPP4i compared with non-East Asians [11]. These results suggested that the effect of DPP4i on the East Asian population is different from other populations.

According to these results, DPP4i affects insulin resistance, however, the mechanisms are not well known in real-world settings, especially in the Asian and Japanese populations. Here, we show that DPP4i improves insulin resistance evaluated by the glucose clamp method in the Japanese population.

# Materials and methods

# Subjects

We recruited 8 Japanese patients with T2DM for this study at Tottori University Hospital from 2013 to 2014. T2DM was diagnosed under the World Health Organization (WHO) criteria [12]. We recruited the patients with age 20–80 years old, HbA1c 6.5-9.0, without oral hypoglycemic agents, and insulin injection. We excluded the patients with cancer, viral hepatitis, liver cirrhosis, pancreatitis, renal failure, or those who are taking medications that increase glucose levels such as corticosteroids. This study was a single-arm study design. The patients took DPP4i as follows; 3 patients with teneligliptin 20 mg, 3 with linagliptin 5 mg, 2 with alogliptin 25 mg. We performed the glucose clamp test and meal tolerance test before and 3 months after DPP4i treatment.

We performed this study according to the Declaration of Helsinki. The Ethics Committee of the Faculty of Medicine, Tottori University approved this study, approval number 2087, approval date 28/01/2013. The first patient was enrolled on 10 April, 2013. The clinical trial number was UMIN000011189, registered (14/07/2013), UMIN000046686, registered (21/01/2022)—Retrospectively registered. We obtained informed consent from all of the participants using a procedure approved by the Ethics Committee.

# Meal tolerance test (MTT)

We conducted MTT as we previously reported [6]. Briefly, the patients ate a test meal developed by the Japan

Diabetes Society (460 kcal; 50% carbohydrate, 35% fat, and 15% protein). We measured plasma glucose, serum insulin at 0, 30, 60, 120, and 180 min after the test meal.

HbA1c was measured by using high-performance liquid chromatography (HPLC). We used HbA1c converter to convert from the percentage values to the mmol/mol values [13].

Insulin resistance and insulin secretion ability were calculated as follows:

 $\label{eq:homeostatic} \begin{array}{lll} Homeostatic & model & assessment & insulin & resistance & (HOMA-IR) & [14] = [fasting & plasma & glucose & (mmol/L)] \times [fasting & plasma & insulin & (pmol/L)]/135. \\ Homeostasis & model & assessment & of & beta-cell & function & (HOMA-beta) & [14] = \{20 \times [fasting & plasma & insulin & (pmol/L)]\}/\{[fasting & plasma & glucose & (mmol/L)] - 3.5\}. \\ \end{array}$ 

Insulinogenic index (IGI) [15] = {[insulin (pmol/L) at 30 min] – [fasting plasma insulin (pmol/L)]}/ $\{[glucose (mmol/L) at 30 min] – [fasting plasma glucose (mmol/L)]\}.$ 

The area under the curve (AUC) was calculated by the horizontal method.

# GLP-1, GIP, and glucagon assays

We obtained blood samples at 0, 30, 60, and 120 min after the meal by using a DPP4 inhibitor-containing blood collection tube (BD<sup>™</sup> P800, BD Japan, Tokyo, Japan). We extracted samples for incretin assay by ethanol and solid-phase method [16]. We measured intact GLP-1 using a GLP-1 (active) ELISA (Merck Millipore, Germany). We measured active GIP using the Human GIP, Active form Assay Kit (IBL, Japan). We measured glucagon levels using a glucagon ELISA (Mercodia, Sweden).

# Hyperinsulinemic-euglycemic clamp

We conducted the glucose clamp test 2 days after MTT as we previously reported [17]. Briefly, we used an artificial endocrine pancreas (STG 55; Nikkiso, Japan), and performed the hyperinsulinemic-euglycemic clamp to measure insulin sensitivity. We used the protocol with a priming insulin infusion at 100 mU/m²/min, and the plasma glucose level was maintained at 5.2 mmol/L (95 mg/dL). This insulin infusion protocol was reported as an insulin level of 1200 pmol/L at steady-state in T2DM patients [18]. The mean GIR during 90–120 min was defined as the glucose disposal rate (GDR), which mainly reflects peripheral insulin sensitivity.

# **Body composition**

Body composition was measured by using Tanita body composition analyzer (Tanita, Japan). We measured the percent body fat, body fat mass, lean body mass, skeletal muscle mass.

# Statistical analysis

We expressed data as the mean  $\pm$  standard deviation. We used the Mann–Whitney U test to assess differences in the mean value of clinical parameters between before and after DPP4i treatment. Values of p < 0.05 were considered to be significant. We used PRISM8 software (GraphPad Software, San Diego, USA) for analyses.

We performed a power analysis to compare GDR before and after treatment as a primary endpoint by using an EZR calculator 23. The mean GDR before treatment was  $4.30\pm1.20$ , and the mean GDR after treatment was  $6.24\pm1.35$ , the difference was  $1.94\pm0.12$ , alpha = 0.05, 1-beta = 0.8, the sample size was 8 patients, the estimated power was 1.0.

# **Results**

Participant characteristics are shown in Table 1. The mean age of patients was  $59.6 \pm 10.4$  years old, and 6 males, 2 females. The duration of T2DM was  $5.8 \pm 6.0$  years. Table 1 shows the effects of DPP4i after 3 months. Bodyweight and HbA1c were significantly changed after 3 months of treatment. Figure 1 shows MTT results. Fasting and postprandial 120 and 180 min glucose were significantly decreased. Insulin levels were not significantly changed. GLP-1 and GIP were significantly increased, but glucagon levels were not changed significantly. The insulin resistance of the glucose clamp was significantly improved, but HOMA-IR was not significantly changed (Table 1). All patients improved insulin resistance derived from the glucose clamp method (data not shown). HOMA-beta, insulinogenic index were not changed. Adiponectin was not changed. Body composition indicated that body fat mass was decreased.

Because the subjects showed significant weight reduction, we calculated GDR  $\times$  BMI/22 to adjust GDR by BMI. The GDR  $\times$  BMI/22 was significantly increased after DPP4i treatment (from  $3.74\pm1.38$  to  $5.73\pm2.16$ , p < 0.001).

# **Discussion**

Our study showed that DPP4i improved the insulin resistance evaluated by the glucose clamp method. DPP4i decreased fasting and postprandial glucose levels without changing insulin levels. It is considered that HOMA-IR generally reflects hepatic insulin resistance, and GDR from glucose clamp reflects muscle insulin resistance [19]. The muscle insulin resistance derived from the glucose clamp method was improved by DPP4i. The HOMA-IR as a marker of liver insulin resistance was not significantly changed, but it was

**Table 1** Comparison between before and after DPP4i treatment

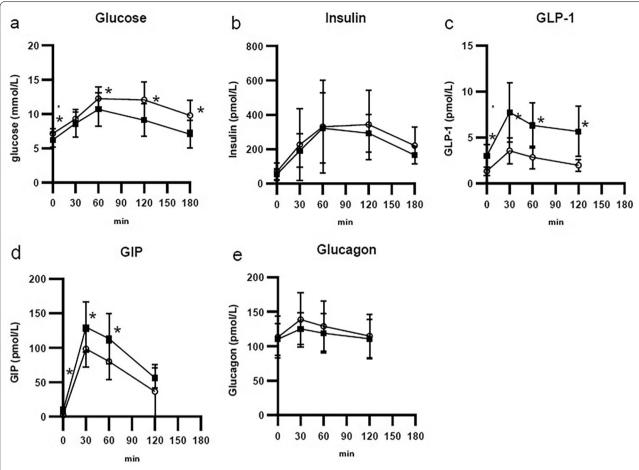
Parameters	Before	After	p value
Body weight (kg)	70.0 ± 12.6	67.8 ± 12.5	< 0.05
BMI (kg/m <sup>2</sup> )	$26.3 \pm 4.8$	$25.3 \pm 4.4$	< 0.05
eGFR	$65.9 \pm 12.8$	$64.2 \pm 13.8$	NS
AST (U/L)	$28.1 \pm 12.7$	$20.5 \pm 3.8$	NS
ALT (U/L)	$43.4 \pm 25.9$	$23.0 \pm 6.8$	< 0.05
Gamma GTP (U/L)	$46.1 \pm 27.4$	$34.5 \pm 11.7$	NS
HbA1c (%)	$7.8 \pm 0.8$	$6.5 \pm 0.7$	< 0.005
HbA1c (mmol/mol)	61±9	$47\pm8$	< 0.005
Insulin secretion			
Fasting Plasma Glucose (mmol/L)	$7.10 \pm 0.70$	$6.22 \pm 1.02$	< 0.05
Glucose AUC (0-120 min)	$21.6 \pm 3.0$	$18.3 \pm 4.1$	< 0.05
Insulin AUC (0-120 min)	$550.3 \pm 387.7$	$496.7 \pm 256.1$	NS
HOMA-beta (%)	$63.8 \pm 42.2$	$79.8 \pm 59.3$	NS
Insulinogenic Index	$1.00 \pm 1.50$	$0.76 \pm 0.68$	NS
Insulin resistance			
HOMA-IR	$3.8 \pm 3.1$	$2.44 \pm 1.19$	NS
GDR (mg/kg/min)	$4.30 \pm 1.2$	$6.24 \pm 1.35$	< 0.01
Incretins			
GLP-1 AUC	$5.2 \pm 1.9$	$12.6 \pm 4.3$	< 0.001
GIP AUC	$128.2 \pm 47.1$	$180.1 \pm 29.3$	< 0.01
Glucagon AUC	$251.9 \pm 67.6$	$234.4 \pm 51.3$	NS
Adiponectin	$5.0 \pm 3.2$	$5.2 \pm 3.2$	NS
Body composition			
Percent body fat (%)	$26.5 \pm 9.0$	$24.4 \pm 7.4$	< 0.05
Body fat mass (kg)	$18.8 \pm 8.9$	$16.9 \pm 7.4$	< 0.05
Lean body mass (kg)	$50.0 \pm 8.8$	$49.8 \pm 8.7$	NS
Skeletal muscle mass (kg)	$47.3 \pm 8.4$	$47.2 \pm 8.3$	NS

Data are presented as the mean  $\pm\,\text{standard}$  deviation

The comparison of parameters between before and after treatment was performed using the Mann–Whitney  $\it U$  test

considered as tending to improvement. Previous studies showed DPP4i increased adiponectin [9], and we previously reported that adiponectin is important for the insulin resistance of Japanese [20]. However, Adiponectin was not changed. There is a report that GLP-1 improved muscle insulin resistance by vasodilatory actions [21]. Our study showed a significant increase of GLP-1, we consider GLP-1 is important for the improvement of muscle insulin resistance. Although further basic research of animals and cellular are needed for understanding the molecular mechanism, our results contribute to understanding the effects of DPP4i.

The previous study also showed DPP4i improves insulin resistance, but this study was performed in American [7]. East Asian and American show different pathophysiology of T2DM, East Asian shows low



**Fig. 1** The meal tolerance test (MTT) results. **a** shows glucose levels after the MTT, **b** insulin, **c** GLP-1, **d** GIP, and **e** glucagon. White circles represent the data from pre-treatment, and black squares indicate post-treatment. Mann–Whitney U test. \*p < 0.05

insulin secretion ability and insulin resistance with mild obesity [8, 22]. East Asians (Japanese, China, Korean, Taiwanese) showed a significantly greater effect of DPP4i compared with non-East Asians [11], our study might contribute to understanding one mechanism of DPP4i. All patients in our study are Japanese, therefore, our results contribute to the daily clinical work in the Japanese population.

Our study did not show a significant change in insulin secretion by DPP4i. Previous studies also did not show a significant change in insulin secretion [4, 7, 9]. A previous study reported that sitagliptin 25 mg and 200 mg increased insulin secretion [5], but this study was performed at 24 h DPP4i treatment, the short period might be different from the real-world situation. A recent large patient number Japanese study showed that DPP4i increased insulin levels, but the increase in insulin levels was very mild [23]. The previous study showed the improvement of insulin resistance without changing insulin levels [7], the authors described that these

findings might be results of improvement of glucose toxicity. We also consider the effects of DPP4i on glucose toxicity is important. Moreover, there are some reports that DPP4i decreases the glucagon section from the alpha cell [24]. We did not see the significant glucagon inhibition by DPP4i, but it may depend on the number of small patients. Furthermore study is needed.

Our study has some limitations, the major limitation is a very small number of patients eight. However, the glucose clamp method and measuring incretins after a meal are very complicated and expensive studies, and it is difficult to recruit patients without any medication in the University hospital. We need a larger study. Our patients showed significant weight reduction. Generally, DPP4i does not change body weight [25], and our study was a single-arm study, we don't have control patients, therefore, our study might have bias. However, the GDR adjusted by BMI also showed a significant increase, we consider DPP4i have the effect of improvement of insulin resistance. However, this BMI

adjusting method might not reflect the true improvement of insulin resistance. We also suspect that body weight/fat reduction in three months directly improved insulin sensitivity, but DPP-4i indirectly through ameliorating glucotoxicity. If we would like to demonstrate the DPP-4i can directly improve insulin sensitivity, we should perform insulin clamp study in short term i.e. one month. Despite these limitations, we consider our study contributes to our daily clinical work.

In conclusion, although the number of patients was very small, these results suggested that DPP4i treatment might improve insulin resistance without changing insulin secretion.

### **Abbreviations**

AUC: Area under the curve; DPP4i: Dipeptidyl peptidase 4 inhibitor; eGFR: Estimate glomerular filtration rate; GDR: Glucose disposal rate; GIP: Glucose-dependent insulinotropic polypeptide; GLP-1: Glucagon-like peptide-1; HbA1c: Glycated hemoglobin; HOMA-beta: Homeostatic model assessment beta-cell function; HOMA-IR: Homeostasis model assessment for insulin resistance.

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### **Author contributions**

All authors contributed significantly. TO participated in the design of the study and performed the statistical analysis. YF, RN, YI, SK, MA, KM, SK, KM, SI, HO, and EU collected the data. MK, TI, ST, and KY conceived the study, participated in its design and coordination, and helped to draft the manuscript. All authors read and approved the final manuscript.

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### Availability of data and materials

The datasets used and/or analysed during the current study available from the corresponding author on reasonable request.

Data sharing statement: No additional data available.

# **Declarations**

# Ethics approval and consent to participate

The Ethics Committee of the Tottori University Faculty of Medicine approved this study (approval number 2087, approval date 28/01/2013). The clinical trial number was UMIN000011189, UMIN000046686. Trial registration: UMIN, UMIN000011189. Registered (14/07/2013)—Retrospectively registered. Trial registration: UMIN, UMIN000046686. Registered (21/01/2022)—Retrospectively registered.

### **Competing interests**

None declared.

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