

RESEARCH

Open Access

Clinical and molecular data from 61 Brazilian cases of Congenital Hyperinsulinemic Hypoglycemia

Raphael Del Roio Liberatore Jr^{1*}, Priscila Manzini Ramos¹, Gil Guerra Jr², Thais Della Manna³, Ivani Novato Silva⁴ and Carlos Eduardo Martinelli Junior¹

Abstract

Objective: To study the clinical and molecular characteristics of a sample of Brazilian patients with Congenital Hyperinsulinemic Hypoglycemia (CHH).

Methods: Electronic message was sent to members from Endocrinology Department- Brazilian Society of Pediatrics requesting clinical data for all cases of CHH. A whole blood sample from living patients was requested for DNA extraction followed by a search for mutations of the genes *ABCC8*, *KCNJ11*, *GCK*, *GLUD1*, *HADH*, *SLC16A1* and *HNF4A*.

Results: Of the 61 patients evaluated, 36 (59%) were boys, and only 16 (26%) were born by normal delivery. Gestational age ranged from 32 to 41 weeks (mean = 37 weeks and 6 days). Birth weight ranged from 1590 to 5250 g (mean = 3430 g). Macrossomia occurred in 14 cases (28%). Age at diagnosis ranged from 1 to 1080 days (mean = 75 days). DNA for molecular analysis was obtained from 53 of the 61 patients. Molecular changes in the *ABCC8* gene were detected in 15 (28%) of these 53 cases, and mutations in the *KCNJ11* gene were detected in 6 (11%). Mutations in the *GLUD1* gene were detected in 9 cases (17%) of the total series. Mutations of the *GCK* gene in heterozygosis were detected in 3 cases. No mutations were detected in the sequencing of genes *HADH*, *SLC16A1* and *HNF4A*.

Conclusion: The present study conducted in Brazil permitted the collaborative compilation of an important number of CHH cases and showed that the present clinical and molecular data are similar to those of published global series.

Keywords: Hypoglycemia, Neonatal, Congenital

Introduction

The maintenance of appropriate blood glucose levels is of vital importance at all ages, but particularly during the neonatal and breastfeeding period. In this phase of life, cerebral tissue is not yet fully mature and is marked by intense metabolic activity and extreme avidity for glucose, being considerably more sensitive to reductions of glycemic levels even if transient. Thus, the occurrence of hypoglycemia in this phase of life is related to high morbidity and mortality [1-4].

Inappropriate insulin secretion in relation to blood glucose levels characterizes hyperinsulinism, which is the main cause of persistent and recurrent hypoglycemia during this phase of life [1,5,6].

The congenital hyperinsulinemic hypoglycemia (CHH) represent a heterogeneous group of clinical condition, the most severe and frequent form of HH [6-8]. The CHH may present as different clinical characteristics (responsive or not to diazoxide, elevated or not ammonia levels) and histological subgroups (focal or diffuse form), but the aspect most recently investigated is the genetic of CHH [1-10].

Mutations in seven different genes (*ABCC8*, *KCNJ11*, *GLUD1*, *GCK*, *HADH*, *SLC16A1* and *HNF4A*) are responsible for about 50% of all CHH cases [1,2]. Of

* Correspondence: rliberatore@fmrp.usp.br

¹Ribeirão Preto Medical School, University of São Paulo, Rua Elzira Sammarco Palma, 400/43, Ribeirão Preto, SP, Brazil

Full list of author information is available at the end of the article

these genes, *ABCC8* and *KCNJ11*, located in regions neighboring chromosome 11 (11p15.1), are responsible for the expression of the proteins SUR1 and Kir6.2. These proteins comprise the potassium channels sensitive to adenosine-triphosphate which play a fundamental role in the control of insulin secretion stimulated by glucose. Inactivating mutations in these two genes are responsible for the most common and severe forms of CHH [9-18]. Defects of the other five genes involved are responsible for a smaller number of cases of CHH [1,2].

Mutations in these genes are expressed with particular histopathological features, clinical manifestations, therapeutic response to drugs and therefore the investigation of these molecular alterations permits a precise diagnosis and an appropriate management.

The objectives of the present study were to obtain clinical information regarding patients with CHH from the largest possible number of clinical centers in Brazil and to determine the molecular etiology of such cases.

Patients and methods

An electronic message was sent to all members of the Department of Endocrinology of the Brazilian Society of Pediatrics, involving representatives of almost all Brazilian states, inviting them to participate in this study. Information about all patients with a diagnosis of CHH was requested, such as sex, gestational age, type of delivery, birth weight and length, age at the onset of symptoms, age at diagnosis of the disease, laboratory results, histological form of the disease, response to drug therapy, need of pancreatectomy, presence of sequelae, and mortality.

The study was approved by the Human Research Ethics Committee of the Faculty of Medicine of Ribeirão Preto, University of São Paulo, and the parents or persons responsible for the living patients gave written informed consent to participate. A whole blood sample was then collected from each index case and his/her parents for DNA extraction followed by the determination of mutations of the seven genes.

DNA was extracted from peripheral blood leukocytes using a commercial kit (QIAamp DNA Mini Kit-Qiagen, Dusseldorf, German) and the sample was stored at -70°C ; the exon ligands of the intron/exons of the genes were amplified by PCR. The PCR product was then sequenced by a standard method using an ABI 3730 sequencer (Applied Bio systems, Warrington, UK) and the sequences obtained were compared to the published sequences [4-9].

Mutations were first searched in the *ABCC8* and *KCNJ11* genes. When no mutations were detected in these genes, mutations were searched in the *GLUD1* and *GCK* genes and in cases of no mutations, the remaining genes were investigated.

Data are reported as absolute and relative frequency, with means and standard deviations.

Results

Data and blood samples of 61 children with CHH were obtained. Blood samples were also obtained from the parents of most of these children, for a total of 140 samples.

Most samples were from the Southeast region of Brazil, which concentrates the largest number of pediatric endocrinology centers. However, the Center-West, Northeast and South regions were also represented. The North region did not send samples.

Of the 61 patients evaluated, 36 (59%) were boys. Gestational age ranged from 32 to 41 weeks (mean = 37 and 6 days; SD = 5 days). Birth weight ranged from 1590 to 5250 g (mean = 3430 g; SD = 480 g). Macrossomia occurred in 14 cases (28%). Birth length was available for 42 patients, but this result was excluded because it was found to be unreliable.

Age at diagnosis ranged from 1 to 1080 days (mean = 75 days; SD = 17 days), with a diagnosis being made at more than 90 days of life in 14 (28%) patients.

The diagnosis of HH are suspect by the need of high rate of glucose infusion to maintain normal blood glucose level, and confirmed by the dosage of low blood glucose level, inadequate insulin level, low ketones and free fatty acids levels in serum [1,2,19]. Other laboratory tests should be collected at the same time with glucose, insulin, ketones and free fatty acids, as ammonia, lactate and glucagon stimulation test for the diagnostic work-out. Except for the glucose and insulin, not all the laboratory tests were carried out for all patients. Blood lactate level was done in 27, ammonia level in 19, ketone level in 17 and free fatty acids in 16 patients. Table 1 lists the results of blood glucose, insulin levels and glucose infusion rate (GIR, mg/kg/min).

The initial drug used as treatment was glucocorticoid in 40 cases (65%). Treatment with diazoxide failure in 24 patients (40%), who were later, submitted to pancreatectomy. All patients submitted to pancreatectomy presented the histological pattern, performed at the different centers, of the diffuse form of the disease.

Table 2 presents the type of treatment performed and the distribution per patient.

DNA quantity and quality sufficient for molecular analysis was obtained from 53 of the 61 patients (87%). In 15 of these 53 cases (28%), molecular changes were detected in the *ABCC8* gene. All mutations were in heterozygosis,

Table 1 Blood glucose, insulin levels and glucose infusion rate (GIR) in 61 patients with CHH

Glucose (mg/dL)	Insulin (IU/mL)	GIR (mg/kg/min)
5-77*	2.5-147*	10-41*
28.5 ± 4.7**	24.9 ± 11.3**	17.2 ± 5.5**

*Range **Median and Standard Deviation.

Table 2 Type of drug used as treatment for 61 patients with CHH

Diazoxide	Glucorticoid	Octreotide	GH*	Nifedipine	Glucagon
41 cases	40 cases	24 cases	14 cases	3 cases	3 cases
67%	66%	39%	23%	5%	5%

*Growth Hormone.

occurring in exon 1 in 6 cases (c.72C > A in 4 and c.134C > T in 2), in exons 31 (c.3992-9G > A) and 36 (c.4415-13G > A) in 2 cases, and in exons 2 (c.257 T > G), 3 (c.331G > A), 4 (c.563A > G), 5 (c.742C > T) and 12 (c.1792C > T) in 1.

Mutations in the *KCNJ11* gene were detected in 6 cases (11%), all of them in heterozygosis, 2 of them being c.801C > G, 2 c.808C > G and 2 c.1142G > A.

The *ABCC8* and *KCNJ11* genes were jointly responsible for mutation in 21 (39%) of 53 cases of CHH.

Analysis of the *GLUD1* gene was carried out in the 32 cases in which no mutations were detected in the *ABCC8* and *KCNJ11* genes. Mutations in this gene were detected in 9 cases (17%), 8 of them being in heterozygosis and 1 in homozygosis. Of these, 7 cases occurred in exon 7 (c.1019A > G), 1 in exon 6 (c.37266C > T) and 1 in exon 10 (c.49322G > T). Serum ammonia was determined in only one of these 9 patients, with a normal result.

Mutations of the *GLUD1* gene were detected in 17% of the total series and in 29% of the cases in which no mutations were detected for the *ABCC8* and *KCNJ11* genes.

The *GCK* gene was analyzed in the 23 cases in which no mutation was detected in the *ABCC8*, *KCNJ11* and *GLUD1* genes. Mutations in heterozygosis were detected in this gene in 3 cases (6%), 2 of them in exon 6 (c.1115C > T) and 1 in exon 10 (c.1829G > T). The *GCK* gene was responsible for 6% of the mutations in the total series and for 10% of the mutations detected in cases that were negative for mutations of the genes associated with the potassium channel.

Sequencing of the *HADH*, *SLC16A1* and *HNF4A* genes did not reveal any mutations.

Table 3 and Figure 1 show the distribution of mutations by gene and also their percentages in the group.

The clinical-molecular correlation with the response to diazoxide and with the histological form could not be determined because in 68% of the cases in which diazoxide was used the dose administered was less than 5 mg/kg/d, not considered ideal for the establishment of the response. All those patients that used low dose, didn't respond to medical treatment.

Discussion

This was the first large-scale study compiling CHH cases in Brazil. Although most samples were from the Southeast

region of the country, all regions were represented, except for the North.

The sample was quite representative in global terms. Recent publications have reported 17 cases for Saudi Arabia [20], 11 cases for China and 33 cases for Italy [21].

The largest samples have been compiled in the two major world reference centers, i.e., England (Great Ormond Hospital) with 300 cases [22] and the United States (Children's Hospital of Philadelphia) with 417 cases [23] and 223 cases of pancreatectomy [24].

As observed in other series, we also could not detect differences regarding gender and prematurity [20-23].

Birth weight ranged from 1590 to 5250 g, with a mean value of 3430 g. In the present series, macrosomia occurred in 14 cases (28%) as compared to 50% of cases in the Italian sample [21]. The diagnosis was made after 90 days of life in 14 (28%) in our children, later than other series, which is a matter of concern since a delayed diagnosis increases the risk of morbi-mortality.

There is still a substantial heterogeneity regarding the diagnostic threshold for the diagnosis of hypoglycemia in Brazil. Regarding the collection of the diagnostic critical sample, there was no standardization of the exams

Table 3 Mutations found in 61 patients with CHH

Gene	Exon	Mutation	Number of patients
ABCC8	1	c.72C > A	4
		c.134C > T	2
	2	c.257 T > G	1
		c.331G > A	1
	4	c.563A > G	1
		c.742C > T	1
	12	c.1792C > T	1
		c.3992-9G > A	2
	36	c.4415-13G > A	2
		KCNJ11	c.801C > G
c.808C > G	2		
2 c.1142G > A.	2		
GLUD1	6	c.37266C > T	1
		c.1019A > G	7
	10	c.49322G > T	1
GCK	6	c.1115C > T	2
		10	c.1829G > T

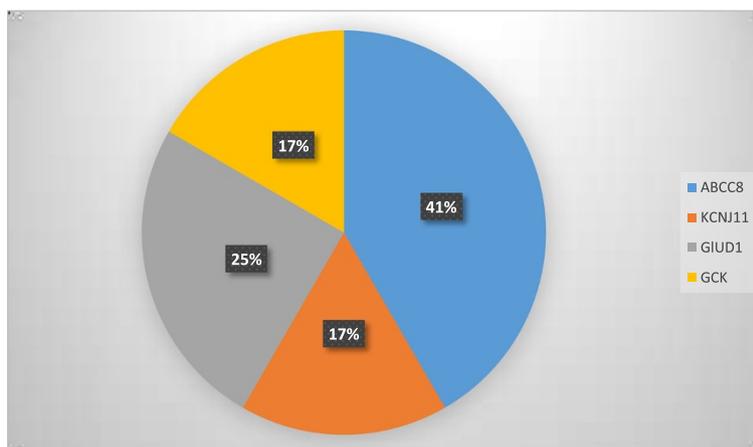


Figure 1 Distribution by gene of the mutations found in 61 patients with CHH.

or of the time when they should be performed. Ammonia was rarely determined, and it should be remembered that hyperammonemia directs the clinical and molecular diagnosis and the clinical conduct regarding the *GLUD1* gene.

There is also a clear need to standardize the treatment, also considering that diazoxide is not a drug easily available in Brazil, requiring importation.

Diazoxide is the medication of choice for the treatment of hypoglycemia due to hyperinsulinism during the neonatal period. None of the other studies have reported the systematic and early use of glucocorticoids, as noted in the present series. Nifedipine is not indicated and the use of growth hormone is indicated only in the presence of its deficiency [25-27].

The lack of response to treatment occurred in 40% of the present cases, a much higher rate than reported in the more recent literature [20-24]. Surgery was possibly indicated before the correct use of drug treatment, with appropriate doses. It was interesting to note that only 2 of the most recent 11 cases required pancreatectomy.

It is noteworthy that 100% of the operated cases had the diffuse form of the disease. According to the literature, 1/3 of the operated cases involve the focal form of the disease [22-24] and this difference requires revision of the histological material and clinical-molecular correlation.

Together, genes *ABCC8* and *KCNJ11* were responsible for mutations in 21 of 53 cases (39%) of CHH. This result is similar to obtained in other series, i.e., 47% of cases in the American series [22] and 40.8% in the British one [23].

A clinical-molecular correlation regarding the response to diazoxide and the histological form was found to be impossible.

Thus, mutations of the *GLUD1* gene were detected in 17% of the total series and in 29% of cases in which

no mutations of the *ABCC8* and *KCNJ11* genes were detected. As reported earlier, this gene is responsible for the clinical form of hypoglycemia associated with high levels of serum ammonia, and is also the major gene affected in cases in which no mutations are detected in genes *ABCC8* and *KCNJ11* [28-31]. Serum ammonia was determined in only one of these patients, with a normal result. A high level of serum ammonia directs the molecular investigation and the drug treatment, as the response to diazoxide is usually exuberant, with no low dose. Unfortunately, diazoxide was not used in these 9 cases, a fact that did not permit us to establish the clinical-molecular relationship reported in the literature [19,28,32-34].

This result has been observed in large series, with the mutations of this gene being as frequent as those of the genes of the potassium channel in the forms responsive to diazoxide [28-31].

The molecular changes in gene *GCK* accounted for 6% of the mutations detected in the entire series and for 10% of the mutations detected in the cases that were negative for mutations in the genes associated with the potassium channel, in agreement with previous literature data [35-38].

In general, mutations were detected in 63% of the series, most of them occurring in the two genes coding for the proteins SUR 1 and Kir6.2, responsible for the functioning of the ATP-dependent potassium channel [39].

Thus, the present pioneering study conducted in Brazil permitted the collaborative compilation of an important number of CHH cases from almost all regions of the country, showing that our clinical and molecular data are similar to those for large global series and underscoring the need to standardize the diagnosis and treatment of CHH in our country.

Abbreviation

CHH: Congenital Hyperinsulinemic Hypoglycemia.

Competing interest

The authors declare that they have no competing interest.

Authors' contributions

RDRL: Author, conceptualized and designed the study, drafted the initial manuscript, and approved the final manuscript as submitted. GGJ: Author; TDM: Author; INS: Author and CEMJ: Author carried out the initial analyses, reviewed and revised the manuscript, and approved the final manuscript as submitted. All authors approved the final manuscript as submitted and agree to be accountable for all aspects of the work.

Funding source

FAPESP Grant 2011/09355-0.

Author details

¹Ribeirão Preto Medical School, University of São Paulo, Rua Elzira Sammarco Palma, 400/43, Ribeirão Preto, SP, Brazil. ²Department of Pediatrics, School of Medical Sciences, State University of Campinas (UNICAMP), Campinas, SP, Brazil. ³Pediatric Endocrine Unit, Instituto da Criança-Hospital das Clínicas, Universidade de São Paulo (USP), São Paulo, SP, Brazil. ⁴Pediatrics Department, Medical School/ Hospital das Clínicas, Universidade Federal de Minas Gerais, Belo Horizonte, Brazil.

Received: 17 November 2014 Accepted: 14 January 2015

Published: 18 February 2015

References

1. Stanley CA. Hyperinsulinism in infants and children. *Pediatr Clin North Am.* 1997;44:363–74.
2. Hussain K, Balnkestein O, De Lonlay P, Christesen HT. Hyperinsulinemic hypoglycemia: biochemical basis and the importance of maintaining normoglycaemia during management. *Arch Dis Child.* 2007;92:56s8–70.
3. Bruninig GJ. Recent advances in hyperinsulinism and the pathogenesis of diabetes mellitus. *Curr Opin Pediatr.* 1990;2:758–65.
4. Fournet JC, Junien C. The genetics of neonatal hyperinsulinism. *Horm Res.* 2003;59:30–4.
5. Glaser B, Thornton PS, Otonkoski T, Junien C. The genetics of neonatal hyperinsulinism. *Arch Dis Child.* 2000;82:79–86.
6. De Lonlay P, Fournet JC, Touati G, Groos MS, Martin D, Sevin C, et al. Heterogeneity of persistent hyperinsulinemic hypo glycaemia. A series of 175 cases. *Eur J Pediatr.* 2002;161:37–48.
7. Meissner T, Mayatepek E. Clinical and genetic heterogeneity in congenital hyperinsulinism. *Eur J Pediatr.* 2002;161:6–20.
8. Sempoux C, Guiot Y, Lefevre A, Nihoul-Fekete C, Jaubert F, Saudubray JM, et al. Neonatal hyperinsulinemic hypoglycemia: heterogeneity of the syndrome and keys for differential diagnosis. *J Clin Endocrinol Metab.* 1998;83:1455–61.
9. Nestorowicz A, Glaser B, Wilson BA, Shyng SL, Nichols CG, Stanley CA, et al. Genetic heterogeneity in familial hyperinsulinism. *Hum Mol Genet.* 1998;7:1119–28.
10. Thomas P, Ye Y, Lightner E. Mutation of the pancreatic islet inward rectifier Kir6.2 also leads to familial persistent hyperinsulinemic hypoglycemia of infancy. *Hum Mol Genet.* 1996;5:1809–12.
11. Thomas PM, Cote GJ, Wohlk N, Haddad B, Mathew PM, Rabl W, et al. Mutations in the sulfonylurea receptor gene in familial persistent hyperinsulinemic hypoglycemia of infancy. *Science.* 1995;268:426–9.
12. Nestorowicz A, Inagaki N, Gono T, Schoor KP, Wilson BA, Glaser B, et al. A nonsense mutation in the inward rectifier potassium channel gene, Kir6.2, is associated with familial hyperinsulinism. *Diabetes.* 1997;46:1743–8.
13. Dunne MJ, Kane C, Shepherd RM, Sanchez JA, James RF, Johnson PR, et al. Familial persistent hyperinsulinemic hypoglycemia of infancy and mutations in the sulfonylurea receptor. *N Engl J Med.* 1997;336:703–6.
14. Otonkoski T, Ammala C, Huopio H, Cote GJ, Chapman J, Cosgrove K, et al. A point mutation inactivating the sulfonylurea receptor causes the severe form of persistent hyperinsulinemic hypoglycemia of infancy in Finland. *Diabetes.* 1999;48:408–15.
15. Tanizawa Y, Matsuda K, Matsuo M, Ohta Y, Ochi N, Adachi M, et al. Genetic analysis of Japanese patients with persistent hyperinsulinemic hypoglycemia of infancy: nucleotide-binding fold-2 mutation impairs cooperative binding of adenine nucleotides to sulfonylurea receptor 1. *Diabetes.* 2000;49:114–20.
16. Marthinet E, Bloc A, Oka Y, Tanizawa Y, Wehrle-Haller B, Bancila V, et al. Severe congenital hyperinsulinism caused by a mutation in the Kir6.2 subunit of the adenosine triphosphate-sensitive potassium channel impairing trafficking and function. *J Clin Endocrinol Metab.* 2005;90:5401–6.
17. Tornovsky S, Crane A, Cosgrove KE, Hussain K, Lavie J, Heyman M, et al. Hyperinsulinism of infancy: novel ABCC8 and KCNJ11 mutations and evidence for additional locus heterogeneity. *J Clin Endocrinol Metab.* 2004;89:6224–34.
18. Ashcroft FM, Harrison DE, Ashcroft SJ. Glucose induces closure of single potassium channels in isolated rat pancreatic beta cells. *Nature.* 1984;312:446–8.
19. Hussain K. Diagnosis and management of Hyperinsulinaemic hypoglycemia of infancy. *Horm Res.* 2008;69:2–13.
20. Al-Agha AE, Ahmad IA. Characterization of the ABCC8 gene mutation and phenotype in patients with congenital hyperinsulinism in western Saudi Arabia. *Saudi Med J.* 2013;34(10):1002–6.
21. SognoValin P, Proverbio MC, Diceglie C, Gessi A, di Candia S, Mariani B, et al. Genetic analysis of Italian patients with congenital hyperinsulinism of infancy. *Horm Res Paediatr.* 2013;79(4):236–42.
22. Kapoor RR, Flanagan SE, Arya VB, Shield JP, Ellard S, Hussain K. Clinical and molecular characterization of 300 patients with congenital hyperinsulinism. *Eur J Endocrinol.* 2013;168(4):557–64.
23. Snider KE, Becker S, Boyajian L, Shyng SL, MacMullen C, Hughes N, et al. Genotype and phenotype correlations in 417 children with congenital hyperinsulinism. *J Clin Endocrinol Metab.* 2013;98(2):E355–63. 98.
24. Lord K, Dzata E, Snider KE, Gallagher PR, De León DD. Clinical presentation and management of children with diffuse and focal hyperinsulinism: A Review of 223 Cases. *J Clin Endocrinol Metab.* 2013;98:E1786–9.
25. Huopio H, Reimann F, Ashfield R, Komulainen J, Lenko HL, Rahier J, et al. Dominantly inherited hyperinsulinism caused by a mutation in the sulfonylurea receptor type 1. *J Clin Invest.* 2000;106:897–906.
26. Thornton PS, MacMullen C, Ganguly A, Ruchelli E, Steinkraus L, Crane A, et al. Clinical and molecular characterization of a dominant form of congenital hyperinsulinism caused by a mutation in the high-affinity sulfonylurea receptor. *Diabetes.* 2003;52:2403–10.
27. Pinney SE, MacMullen C, Becker S, Lin YW, Hanna C, Thornton P, et al. Clinical characteristics and biochemical mechanisms of congenital hyperinsulinism associated with dominant KATP channel mutations. *J Clin Invest.* 2008;118:2877–86.
28. Stanley CA, Fang J, Kutyna K, Hsu BY, Ming JE, Glaser B, et al. Molecular basis and characterization of the hyperinsulinism/hyperammonemia syndrome: predominance of mutations in exons 11 and 12 of the glutamate dehydrogenase gene. *Diabetes.* 2000;49:667–73.
29. Yorifuji T, Muroi J, Uematsu A, Hiramatsu H, Momoi T. Hyperinsulinism hyperammonemia syndrome caused by mutant glutamate dehydrogenase accompanied by novel enzyme kinetics. *Hum Genet.* 1999;104:476–9.
30. Santer R, Kinner M, Passarge M, Superti-Furga A, Mayatepek E, Meissner T, et al. Novel missense mutations outside the allosteric domain of glutamate dehydrogenase are prevalent in European patients with the congenital hyperinsulinism-hyperammonemia syndrome. *Hum Genet.* 2001;108:66–71.
31. Fujioka H, Okano Y, Inada H, Asada M, Kawamura T, Hase Y, et al. Molecular characterization of glutamate dehydrogenase gene defects in Japanese patients with congenital hyperinsulinism/hyperammonemia. *Eur J Hum Genet.* 2001;9:931–7.
32. James C, Kapoor RR, Ismail D, Hussain K. The genetic basis of congenital hyperinsulinism. *J Med Genet.* 2009;46:289–99.
33. Kapoor RR, James C, Hussain K. Advances in the diagnosis and management of Hyperinsulinemic hypoglycemia. *Nature Clin Pract Endomet.* 2009;5:101–12.
34. Kapoor RR, Flanagan SE, James C, Shield JP, Ellard S, Hussain K. Hyperinsulinaemic hypoglycemia. *Arch Dis Child.* 2009;94:450–7.
35. Glaser B, Kesavan P, Heyman M, Davis E, Cuesta A, Buchs A, et al. Familial hyperinsulinism caused by an activating glucokinase mutation. *N Engl J Med.* 1998;338:226–30.
36. Gloyn AL, Noordam K, Willemsen MA, Ellard S, Lam WW, Campbell IW, et al. Insights into the biochemical and genetic basis of glucokinase activation from naturally occurring hypoglycemia mutations. *Diabetes.* 2003;52:2433–40.

37. Wabitsch M, Lahr G, Van de Bunt M, Marchant C, Lindner M, von Puttkamer J, et al. Heterogeneity in disease severity in a family with a novel G68V GCK activating mutation causing persistent hyperinsulinemic hypoglycemia of infancy. *Diabet Med.* 2007;24:1393–9.
38. Christesen HB, Tribble ND, Molven A, Siddiqui J, Sandal T, Brusgaard K, et al. Activating glucokinase (GCK) mutations as a cause of medically responsive congenital hyperinsulinism: prevalence in children and characterization of a novel GCK mutation. *Eur J Endocrinol.* 2008;159:27–34.
39. Liberatore Jr RDR, Martinelli Jr CE. Hypoglycemia hyperinsulinemic of infancy. *Arq Bras Endocrinol Metab.* 2011;55(3):177–83.

doi:10.1186/1758-5996-7-5

Cite this article as: Del Roio Liberatore *et al.*: Clinical and molecular data from 61 Brazilian cases of Congenital Hyperinsulinemic Hypoglycemia. *Diabetology & Metabolic Syndrome* 2015 **7**:5.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

