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ADIPOQ and *IL6* variants are associated with a pro-inflammatory status in obesities with cardiometabolic dysfunction

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Abstract

Background: Polymorphisms in genes encoding adiponectin (*ADIPOQ*) and interleukin-6 (*IL6*) have been associated with adiposity and obese-related phenotypes. This study investigated the relationship of *ADIPOQ* and *IL6* gene polymorphisms with pro-inflammatory and cardiometabolic markers in obese patients.

Methods: Anthropometric and body composition parameters were measured in 249 Brazilian subjects (30 to 68 yr). Metabolic and inflammatory markers and adipokines were analyzed in blood samples. *ADIPOQ* rs2241766 (45 T > G) and *IL6* rs1800795 (-174G > C) polymorphisms were analyzed by real-time PCR and PCR-RFLP, respectively.

Results: Type 2 diabetes, hypertension, dyslipidemia and increased values of waist circumference, body fat, leptin, fibrinogen, IL-1 β , hsCRP and TNF α were related to obesity ($p < 0.05$). Multiple linear regression analysis showed a positive correlation between BMI and waist circumference, body fat, leptin, fibrinogen, PAI-1, IL-1 β , hsCRP and TNF α values ($p < 0.001$) but not with adiponectin. Obese group had altered metabolic status, resistance to leptin and insulin, and atherogenic and pro-inflammatory profiles. *ADIPOQ* and *IL6* variants were not directly related to obesity, leptin resistance or alterations in cardiometabolic markers. Individuals carrying *ADIPOQ* 45G allele (TG + GG genotype) had higher IL-6, IL-1 β and TNF α levels than TT genotype carriers ($p < 0.05$). *IL6* -174GG genotype was associated with increased IL-1 β levels ($p = 0.033$).

Conclusion: Obesity is associated with leptin resistance, cardiometabolic alterations and a pro-inflammatory status. Our results are suggestive that *ADIPOQ* and *IL6* polymorphisms contribute to cardiometabolic risk in obese individuals.

Keywords: Obesity, Inflammation, Adiponectin, Interleukin-6, Gene polymorphism

Background

Obesity is a multifactorial clinical condition caused by excessive adiposity that is a major contributor to the global epidemic of type 2 diabetes (T2DM), fatty liver disease and cardiovascular disease (CVD) [1,2].

Adipose tissue is a key endocrine organ, which produces several bioactive molecules (adipokines), such as leptin, resistin and adiponectin, with pro- or anti-inflammatory activities [3]. Adipokines are involved in the regulation of appetite and satiety, energy expenditure, endothelial function, hemostasis, blood pressure, adipogenesis and fat

distribution, and insulin secretion and sensitivity [4]. Therefore, expansion of adipose tissue changes the secretion of adipokines towards a pro-inflammatory, diabetogenic and atherogenic pattern [3-5].

Adiponectin is an adipokine that is involved in the sensitivity to insulin. Adiponectinemia is markedly decreased in individuals with visceral obesity and states of insulin resistance, such as nonalcoholic fatty liver disease, atherosclerosis, and T2DM [6]. Therefore, adiponectin has been considered as a biomarker for insulin resistance, T2DM, metabolic syndrome and cardiovascular disease.

Adipose tissue also secretes several inflammatory molecules, including interleukin-6 (IL-6) that has both local and systemic effects. It has been suggested that IL-6

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plays a role in the induction of the inflammatory process linked to obesity [7].

Common single nucleotide polymorphisms (SNPs) in adiponectin (*ADIPOQ*) and IL-6 (*IL6*) genes have been shown to be related to obesity and adiposity-related phenotypes.

The *ADIPOQ* rs2241766 (45 T > G) and rs1501299 (276G > T, c.214 + 62G > T) SNPs have been associated with variability in plasma adiponectin levels [8,9]. Moreover, these variants have been also related to increased body weight, adiposity, metabolic alterations, insulin resistance, and risk for T2DM and coronary artery disease (CAD), in studies from different ethnic populations [10-15].

A common SNP located in the promoter region of *IL6*, known as -174G > C (rs1800795, c.-237C > G) variant was associated with increased risk for overweight and obesity-related metabolic disorders, especially insulin resistance in various group populations [16-19]. This *IL6* variant was suggested to play a role in the regulation of body mass through their influence on metabolism and energetic homeostasis [20].

We have investigated the relationship between *ADIPOQ* 45 T > G and *IL6* -174G > C variants and obesity and metabolic markers in a sample of our population.

Subjects and methods

Study subjects

Two-hundred-forty-nine individuals, aged 30 to 68 yr, were randomly selected at the University Hospital of University of Sao Paulo, Sao Paulo, Brazil. They were informed about the study protocol and those who agreed to participate as volunteers have signed the informed consent. The study protocol was approved by the Ethics Committees of the School of Pharmaceutical Sciences and University Hospital of the University of Sao Paulo (Sao Paulo, Brazil).

Individuals with thyroid, adrenal or gonad dysfunction or other type of secondary obesity, as well as liver, kidney or gastrointestinal disease, and pregnant women or under treatment of oral contraceptives were not included in the study.

All participants answered a questionnaire to collect personal information during an interview. Each individual declared his ethnic group and provide information about medication use, physical activity, alcohol consumption and cigarette smoking.

Anthropometric measurements, such as body mass index (BMI), waist circumference, waist-to-hip ratio (WHR) were taken from each participant. Individuals were grouped as normoweight (BMI \leq 24.9 kg/m²), overweight (BMI 25.0-29.9 kg/m²) and obese (BMI \geq 30 kg/m²). Body fat and basal metabolic rate (BMR) were measured using a 310E impedanciometer (RJL Systems, Inc, USA).

Systolic/diastolic blood pressure was measured in supine position after resting for 30 min by a trained physician using a mercury column sphygmomanometer. Subjects with systolic/diastolic blood pressure over to 140/90 mmHg or were under anti-hypertensive therapy were considered hypertensive.

Participants with fasting glycemia over 125 mg/dL (American Diabetes Association), or taking a glucose-lowering drug were classified as type 2 diabetics. Current tobacco smoking was considered as a daily intake of one or more cigarette. Alcohol consumption was considered an intake of any dose of beer, wine and/or distilled spirits according to World Health Organization recommendation. Physical exercise practice was considered the practice of sports, for example walking, running or swimming, for at least 2 h per week.

Laboratory tests

Blood samples were drawn for genomic DNA extraction and measurements of metabolic and inflammatory markers after fasting for 12 h. Serum glucose, total cholesterol, triglycerides and high density lipoprotein (HDL) cholesterol were measured by enzyme-colorimetric methods and apolipoprotein (apo) AI and B were determined by immunoturbidimetry (Siemens Healthcare Diagnosis Inc., Tarrytown, NY, USA). Values of very-low (VLDL) and low density lipoprotein (LDL) cholesterol were calculated.

Fibrinogen was determined using the Fibre Kit-Test Diagnostica Stago Inc (Bayer/STACompact-Stago, Germany). High sensitive C reactive protein (hsCRP) and insulin were determined by immunonephelometry and chemiluminescence, respectively (Siemens Healthcare Diagnosis Inc., Tarrytown, NY, USA). HbA1c was measured in EDTA-anticoagulated blood by affinity chromatography (HPLC) using the D10 Hemoglobin Testing System (Biorad [®], San Francisco, USA). The homeostasis model assessment (HOMA) was used for evaluation of the beta-cell secretory function (HOMA- β) and insulin resistance (HOMA-IR).

Leptin, adiponectin, tumor necrosis factor alpha (TNF α), IL-6, interleukin 1beta (IL-1 β), and plasminogen activator inhibitor 1 (PAI-1) in serum were measured by Milliplex technology Luminex Map [®] 100/200 (Gen-Probe, Austin, TX, USA). Soluble leptin receptor (sLEPR) was measured by ELISA using kit from ALEXIS Biochemicals Axxora, LLC (San Diego, CA, USA).

Genetic analysis

Genomic DNA was extracted from peripheral blood leukocytes using the method of affinity chromatography system using QIAGEN spin-column kits and robotic workstation to purify DNA QIAcube (Qiagen Biotechnology Brasil Ltda, Sao Paulo, Brazil).

The *ADIPOQ* 45 T > G (rs2241766) SNP was genotyped by TaqMan real time PCR using the pre-designed assay ID c__26426077_10 for allelic discrimination, containing specific probes for each allele marked with VIC and FAM fluorescent dyes (Applied Biosystems, Foster City, USA). The real time PCR was performed in a ABI PRISM 7500 FAST (Applied Biosystems, Foster City, CA, USA) using the following program: a start cycle of 2 min at 50°C, one cycle of 10 min at 95°C, and 40 cycles of 15 s at 95°C and 1 min at 60°C. The fluorescence signals were analyzed using the program Sequence Detection Software V 1.2.3 (Applied Biosystems, Foster City, USA) that generates clusters of signal amplification which allow the identification of each genotype. Samples with each genotype were analyzed together as an internal control.

The *IL6* -174G > C (rs1800795) SNP was detected by PCR-RFLP. The PCR primers were designed based on the *IL6* sequence [NM_000600] (GenBank, NIH/NCBI) using the Primer Premier® v.5.0 software (PremierBiosoft International, USA), as follows: forward, 5'-TGACTT-CAGCTTTACTCTTTGT-3'; and reverse, 5'-CTGAT TGGAAACCTTATTAA-3'. The genotypes were identified by an end point PCR assay, using 50 ng DNA, 200 nmol/L primers (IDT, Coralville, IA, USA), 200 μmol/L dNTPs (GE Healthcare, Amersham Biosciences do Brasil, São Paulo, Brazil), 1 U DNA polymerase (BioTools, Madrid, Spain), PCR buffer [75 mM Tris-HCl (pH 9.0), 50 mM KCl, 20 mM (NH₄)₂SO₄, 2 mmol/L MgCl₂] in 50 μL. PCR assays were carried out in a Mastercycler® (Eppendorf AG, Hamburg, Germany) using the following program: a cycle of 95°C for 1 min, 30 cycles of 95°C for 1 min, 59°C for 1 min and 72°C for 1 min; and a cycle of 72°C for 10 min. The 226pb-amplicon was digested with the endonuclease *Nla* III according to the manufacturer (New England Biolabs Inc., Ipswich, MA, USA). Restriction fragments were identified by 2% agarose gel electrophoresis.

DNA genotyping results were interpreted by two independent analysts and 20% of DNA samples were randomly re-analyzed.

Statistical analysis

The results were analyzed using the SigmaStat v. 2.03 (Systat software Inc., San Jose, CA, USA) and Minitab v.15 statistical software (Minitab Inc. State College, PA, USA), assuming significance level of $p < 0.05$. Chi-square test was used to compare categorical variables and the agreement of genotypes frequencies with Hardy-Weinberg equilibrium (HWE) expectations. Continuous variables were compared by *t*-test or Mann-Whitney Rank sum test, as well as ANOVA and Kruskal Wallis with all pairwise multiple comparison using Holm-Sidak and Dunn methods, respectively. Multiple linear regression

analysis was used to establish correlations between body mass index and independent variables, considering the covariates: age, gender, ethnics, hypertension, tobacco smoking, alcohol consumption, physical exercise practice, T2DM and CAD. Nominal logistic regression analysis was performed to evaluate the influence of clinical, biochemical and genetic parameters on the risk for overweight and obesity, using the normoweith as reference group.

Results

Table 1 shows clinical, anthropometrics and laboratory data of the individuals enrolled in this study. Female was the predominant gender in this sample, with higher proportion in normoweight than in overweight and obese groups ($p = 0.035$). On the other hand, menopause, hypertension, T2DM was less frequent in normoweight individuals ($p < 0.05$), while obese group was less prone to practice of physical activity ($p = 0.003$). Mean age, ethnics, family history of CAD, tobacco smoking and alcohol consumption had similar values among the studied groups ($p > 0.05$). As expected, measurements of BMI, waist circumference, WHR, body fat content and BMR were higher in the obese group and lower in normoweight group ($p < 0.001$). The leptinemia was higher and sLEPR levels were lower in obese compared with those from normoweight individuals ($p < 0.05$), while no differences were found in adiponectin plasma concentration among the groups ($p > 0.05$).

Obese subjects had higher levels of glucose, HbA1c, HOMA-β and HOMA-IR than normoweight group ($p < 0.05$). Obese also had a more atherogenic profile with higher levels of LDL and VLDL cholesterol, triglycerides and apoB and lower HDL cholesterol and apoAI compared with the normoweight group ($p < 0.05$). The values of fibrinogen, PAI-1, IL-6, IL-1β, TNFα and hsCRP were also higher in obese than in normoweight subjects ($p < 0.05$) (Table 1).

Results from multiple linear regression analysis confirmed that waist circumference, WHR and body fat were positively correlated with BMI values ($p < 0.001$) (Table 2). In addition, an increment of one unit in the plasma concentration of leptin, IL-1β, hsCRP, TNFα, PAI-1 and fibrinogen were related respectively with an increase of 0.12, 0.11, 0.31, 0.35, 0.12, 0.02 Kg/m² of BMI in the study group ($p < 0.05$).

Univariate logistic regression analysis was carried out to evaluate the influence of clinical, biochemical and genetic variables as risk factors for overweight and obesity. As shown in Table 3, waist circumference, body fat and dyslipidemia increased the risk for overweight and obesity in this sample ($p < 0.05$). In addition, hypertension and T2DM increase more than four times the risk for obesity ($p < 0.001$), while female gender and physical activity reduce this risk ($p < 0.05$). Increased plasma

Table 1 Clinical, anthropometrics, laboratory and polymorphism data of the studied group

Variable	Normoweight (51)	Overweight (53)	Obese (145)	P-value
Age, years	46.6 ± 8.3	47.9 ± 6.9	48.7 ± 9.2	0.323
Ethnicity [White], %	78.4 (40)	69.8 (37)	71.0(103)	0.539
Woman, %	92.2 (47)	79.2 (42)	75.2 (109)	0.035
Menopause,%	10.6 (5)	26.2 (11)	33.9 (37)	0.011
Hypertension, %	13.7 (7)	28.3 (15)	45.5 (66)	<0.001
Type 2 diabetes, %	33.3 (17)	41.5 (22)	55.7 (99)	<0.001
Family history of CAD	11.7 (6)	20.8 (11)	17.9 (26)	0.455
Tobacco smoking,%	17.6 (9)	23.2 (10)	13.1 (19)	0.527
Alcohol consumption, %	2.0 (1)	3.8 (2)	3.4 (5)	0.845
Physical exercise practice,%	50.9 (26)	50.9 (27)	29.6 (43)	0.003
Body mass index, kg/m ²	22.6 ± 1.7a	27.4 ± 1.4b	35.1 ± 4.2c	<0.001
Waist circumference, cm	73.9 ± 8.7a	86.8 ± 9.4b	104.1 ± 11.0c	<0.001
Waist- hip ratio	0.78 ± 0.07a	0.84 ± 0.08b	0.90 ± 0.10 c	<0.001
Body fat, %	30.9 ± 4.7a	33.9 ± 6.4b	38.5 ± 5.0c	<0.001
BMR, kcal	1272 ± 240a	1501 ± 295b	1742 ± 280c	<0.001
Leptin, ng/mL	13.6 ± 8.1a	15.7 ± 12.0a,b	22.9 ± 19.2b	0.012
sLEPR, ng/mL	23.8 ± 12.4a	19.1 ± 13.5a,b	20.0 ± 24.0b	0.011
Adiponectin, µg/mL	24.6 ± 23.2	17.7 ± 23.0	27.2 ± 26.4	0.070
Glucose, mg/dL	92 ± 9a	102 ± 26a,b	106 ± 29b	<0.001
HbA1c, %	5.5 ± 0.4a	6.0 ± 1.3a,b	6.0 ± 1.4b	0.005
Insulin, mU/L	6.6 ± 4.3a	9.7 ± 6.1b	21.3 ± 12.2b	<0.001
HOMA-β	23.0 ± 18.2a	33.3 ± 24.5b	68.7 ± 37.2b	<0.001
HOMA-IR	1.5 ± 1.0a	2.4 ± 1.5b	5.9 ± 4.8b	<0.001
Total cholesterol, mg/dL	194 ± 36	203 ± 37	209 ± 40	0.077
HDL cholesterol, mg/dL	65 ± 19a	55 ± 17b	51 ± 12b	<0.001
LDL cholesterol, mg/dL	113 ± 32a	125 ± 32a,b	128 ± 32b	0.016
VLDL cholesterol, mg/dL	16 ± 8a	23 ± 10b	30 ± 18c	<0.001
Triglycerids, mg/dL	79 ± 38a	114 ± 53b	150 ± 88c	<0.001
Apolipoprotein A1, mg/dL	160 ± 40a	152 ± 36a,b	146 ± 31b	0.009
Apolipoprotein B, mg/dL	89 ± 28a	109 ± 38b	101 ± 28b	0.004
Fibrinogen, mg/dL	341 (304–374)a	350 (319–409)a,b	374 (328–446)b	<0.001
PAI-1, ng/mL	62.5 (44.7–76.1)a	63.6 (51.0–95.4)a	96.1(60.0–150.6)b	<0.001
IL-6, pg/mL	0.08 (0.07–0.09)a	0.09 (0.08–0.55)a,b	0.56 (0.28–1.6)c	<0.001
IL-1β, pg/dL	5.0 (4.0–5.6)a	5.9 (4.0–6.1)a,b	18.0 (15.0–51.5)c	<0.001
hsCRP, mg/L	0.14 (0.05–0.58)a	0.49 (0.08–1.17)a,b	1.72 (0.26–5.69)c	<0.001
TNFα, pg/mL	0.56 (0.09–1.25)a	0.71 (0.32–2.85)a,b	3.17 (1.45–6.05)c	<0.001
Minor Allele Frequency, %				
ADIPOQ 45 T > G (rs2241766)	19.6	22.6	21.7	0.858
IL6 -174G > C (rs1800795)	26.4	37.7	31.0	0.209

Number of individuals is in parenthesis. Results are shown as mean ± SD or median (interquartile range) and compared by Anova (Multiple comparisons by Holm-Sidak method) or Kruskal Wallis (Dunn's Method). Different letters indicate differences between mean values. Categorical variables were compared by chi-square. BMR: basal metabolic rate; CAD: coronary artery disease; HDL: high-density lipoprotein; hsCRP: high sensitive C reactive protein; LDL: low-density lipoprotein; PAI-1: plasminogen activator inhibitor-1; IL-6: interleukin 6; IL-1β: interleukin 1beta; sLEPR: soluble leptin receptor; TNFα: tumor necrosis factor alpha; VLDL: very low-density lipoprotein.

Table 2 Anthropometric, biochemical and genetic variables as predictors for body mass index values: multiple linear regression analysis

Independent variables	B	SE	P-value
<i>Anthropometrics</i>			
Waist circumference, cm	0.361	0.017	<0.001
Body fat, %	0.840	0.055	<0.001
Waist-hip ratio	21.317	4.397	<0.001
<i>Biochemical parameters</i>			
Fibrinogen, mg/dL	0.021	0.004	<0.001
PAI-1, pg/mL	0.121	0.038	0.002
hsCRP, mg/L	0.31	0.07	<0.001
IL-6, pg/mL	-0.006	0.118	0.962
IL-1 β , pg/dL	0.110	0.018	<0.001
TNF α , pg/mL	0.348	0.130	0.008
Adiponectin, μ g/mL	0.0023	0.0170	0.890
Leptin, ng/mL	0.120	0.024	<0.001
sLEPR, ng/mL	0.0007	0.0232	0.975
<i>Polymorphisms</i>			
<i>ADIPOQ</i> 45 T > G (ref TT)	0.259	0.773	0.738
<i>IL6</i> -174G > C (ref GG)	0.099	0.740	0.893

Regression coefficients (B) and standard errors (SE) are expressed in kg/m². Age, gender, ethnics, hypertension, tobacco smoking, alcohol consumption, physical exercise practice, T2DM and CAD were used as covariates. Polymorphisms were introduced as dummy variables for absence or presence of the rare allele.

levels of both fibrinogen and IL-1 β were associated with high risk for overweight and obesity ($p < 0.05$) (Table 3). Moreover, increased leptin, hsCRP and TNF α concentrations were also risk factors for obesity ($p < 0.05$).

ADIPOQ 45 T > G and *IL6* -174G > C SNPs had genotype distributions as expected from HWE in this sample (Data not shown). Minor allele frequencies of these variants were similar between obese, overweight and normo-weight groups ($p > 0.05$) (Table 1). These variants did not show significant association with neither BMI (Table 2) nor risk for overweight and obesity (table 3) in this sample. Interestingly, individuals carrying *ADIPOQ* 45G allele (TG + GG genotype) had higher IL-6, IL-1 β and TNF α plasma levels than TT genotype carriers ($p < 0.05$) (Figure 1, Table 4). While *IL6* -174GG genotype was associated with higher levels of IL-1 β ($p = 0.033$) (Figure 1, Table 5).

Discussion

Results from this study demonstrate an association of obesity with hypertension, T2DM, insulin resistance and an atherogenic lipid profile, confirming that overweight and obese individuals are more susceptible to metabolic dysfunction and atherosclerosis, which are known risk factors for CVD [21].

The hyperleptinemia and reduced sLEPR found in overweight and obese patients indicate a status of leptin resistance, which is probably caused by dysregulation of the negative feedback, as a classic mechanism of hormone resistance [22]. It is well known that hyperleptinemia triggers a chronic overstimulation of the leptin receptor and activation of negative feedback pathways that block further leptin signaling, leading to leptin resistance [22]. The hyperleptinemia has been attributed to a deficiency in leptin transport through the blood-brain barrier, as well as, to the presence of variants in *LEPR*, which alters functionally or even expression of the receptor reducing its circulating levels, as found in obese subjects.

It has been suggested that resistance to leptin produce metabolic and inflammatory alterations in several tissues and organs, including the liver, spleen and heart, therefore leptin resistance contributes to the risk for obesity-related comorbidities [23].

In this work, obesity was associated with hyperglycemia, insulin resistance and dyslipidemia. Obese patients have reduced supply of glucose in adipocytes that leads to a decreased intracellular lipolysis and increased release of non-esterified fatty acids, which results in insulin resistance, dysglycemia and dyslipidemia [24]. These metabolic alterations increase the risk for CVD in obese subjects.

We could not find a direct relationship of the *ADIPOQ* 45 T > G polymorphism with obesity or variability in body fat mass, waist circumference and BMI values. Similarly, this variant was not associated with BMI in other groups such as Mexican-Mestizos [25], Tunisian volunteers [26], Saudi Arabians [27] and Chinese [28]. This lack of association was also found in groups of women [29] or men [10,11] suggesting that is independent on gender.

The *ADIPOQ* 45 T > G variant has been shown to be associated with variability in adiponectin levels in several studies including in overweight Finish subjects with impaired glucose tolerance [12], diabetic and non-diabetic Brazilian patients with high cardiovascular risk [15], Arab patients with acute coronary syndrome [30]. However, this relationship was not found in our study and in other population samples [25-29], including patients with coronary artery disease [31,32].

We did not find an association between *IL6* -174G > C SNP and obesity or variability in BMI. A large study with two independent cohorts has indicated that *IL6* variants are significantly associated with adiposity, but the contribution of the SNP -174G > C seems to be less likely [33]. Lack of relationship of this variant and obesity was also found in children [34], adolescents [35] and diabetic adults [36] suggesting that it is not a major contributor to obesity risk.

Table 3 Influence of clinical, biochemical and genetic variables on the risk for overweight and obesity: Univariate logistic regression analysis

Independent variables	Risk for overweight			Risk for Obesity		
	OR	95% CI	P-value	OR	95% CI	P-value
<i>Clinical and anthropometric variables</i>						
Age, years	1.02	0.97 – 1.06	0.453	1.03	0.99 – 1.07	0.145
Gender [ref: male]	0.37	0.11 – 1.25	0.110	0.25	0.08 – 0.73	0.011
Ethnics [ref: white]	0.64	0.26 – 1.55	0.318	0.67	0.31 – 1.43	0.297
Menopause	1.13	0.50 – 2.52	0.774	0.54	0.27 – 1.08	0.081
Hypertension	2.48	0.92 – 6.72	0.074	5.32	2.25 – 12.60	<0.001
T2DM	1.42	0.64 – 3.15	0.390	4.40	2.23 – 8.69	<0.001
Dyslipidemia	3.30	1.48 – 7.38	0.004	5.02	2.53 – 9.97	<0.001
Family history of CAD						
Tobacco smoking	1.09	0.40 – 2.94	0.872	0.71	0.30 – 1.69	0.437
Alcohol consumption	1.96	0.17 – 22.32	0.587	1.80	0.21 – 15.77	0.596
Physical exercise practice	0.93	0.43 – 2.00	0.845	0.41	0.21 – 0.79	0.008
Waist circumference, cm	1.21	1.12 – 1.30	<0.001	1.42	1.30 – 1.55	<0.001
Body fat, %	1.13	1.04 – 1.23	0.004	1.34	1.23 – 1.46	<0.001
<i>Biochemical parameters</i>						
Leptin, ng/mL	1.02	0.98 – 1.06	0.434	1.05	1.01 – 1.08	0.006
sLEPR, ng/mL	0.99	0.90 – 1.01	0.397	0.99	0.97 – 1.01	0.421
Adiponectin, µg/mL	0.99	0.97 – 1.01	0.155	1.00	0.99 – 1.02	0.555
Fibrinogen, mg/dL	1.01	1.00 – 1.01	0.037	1.01	1.00 – 1.01	<0.001
PAI-1, pg/mL	1.03	0.98 – 1.08	0.204	1.04	1.00 – 1.08	0.057
IL-6, pg/mL	1.23	0.91 – 1.67	0.183	1.24	0.92 – 1.66	0.154
IL-1β, pg/dL	1.08	1.01 – 1.16	0.029	1.14	1.06 – 1.22	<0.001
hsCRP, mg/L	1.24	0.87 – 1.76	0.232	1.75	1.28 – 2.3	<0.001
TNFα, pg/mL	1.18	0.96 – 1.45	0.123	1.35	1.13 – 1.62	0.001
<i>Polymorphisms</i>						
ADIPOQ 45 T > G (ref TT)	1.03	0.46 – 2.32	0.946	1.06	0.54 – 2.09	0.858
IL6 -174G > C (ref GG)	1.50	0.68 – 3.30	0.317	0.95	0.50 – 1.81	0.884

Results obtained from a univariate nominal regression analysis using the normoweight subjects as reference group. OR, odds ratio; CI, confidence interval. Polymorphisms were introduced as dummy variables for absence or presence of the rare allele.

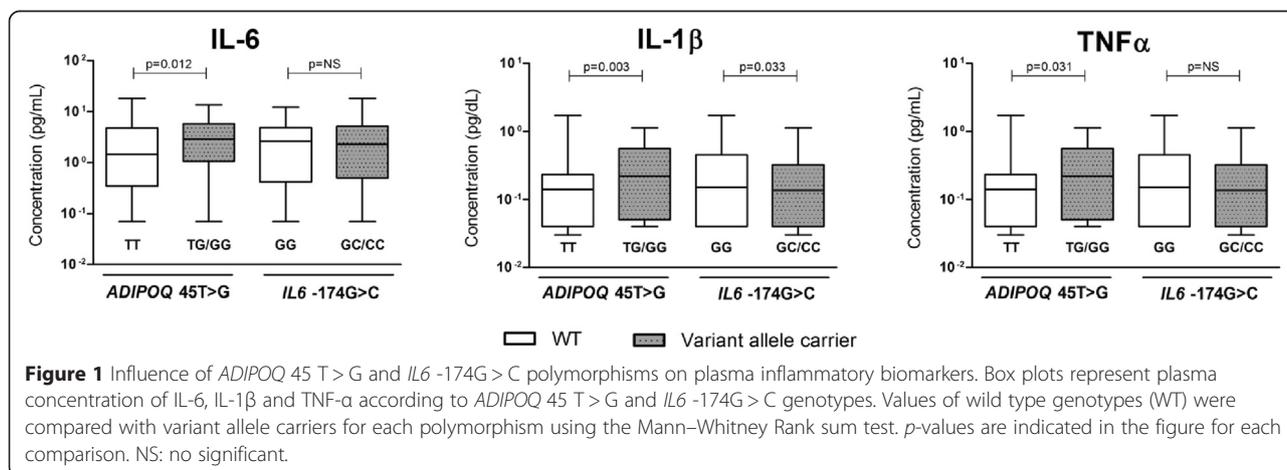


Table 4 Relationship of *ADIPOQ* 45 T > G with anthropometric, metabolic and inflammatory variables

Variable	Genotypes		P-value
	TT (163)	TG + GG (86)	
Body mass index, kg/m ²	30.9 ± 6.2	31.1 ± 6.2b	0.818
Waist circumference, cm	94.4 ± 16.4	94.1 ± 15.3	0.881
Waist- hip ratio	0.86 ± 0.10	0.87 ± 0.10	0.249
Body fat, %	35.8 ± 6.0	36.3 ± 6.5	0.527
BMR, kcal	1611 ± 336	1530 ± 328	0.190
Leptin, ng/mL	20.5 ± 17.6	18.8 ± 15.8	0.666
sLEPR, ng/mL	21.0 ± 21.9	19.6 ± 18.6	0.614
Adiponectin, µg/mL	25.7 ± 25.4	21.5 ± 24.0	0.442
Glucose, mg/dL	101 ± 25	104 ± 27	0.464
HbA1c, %	5.9 ± 1.2	5.9 ± 1.3	0.476
Insulin, mU/L	15.3 ± 1.6	17.6 ± 12.7	0.181
HOMA-β	50.7 ± 38.6	56.0 ± 35.3	0.205
HOMA-IR	4.1 ± 4.0	5.0 ± 4.8	0.153
Total cholesterol, mg/dL	205 ± 36	204 ± 43	0.663
HDL cholesterol, mg/dL	55 ± 15	53 ± 16	0.205
LDL cholesterol, mg/dL	125 ± 32	124 ± 34	0.816
VLDL cholesterol, mg/dL	25 ± 14	27 ± 19	0.475
Triglycerides, mg/dL	124 ± 70	135 ± 93	0.475
Apolipoprotein AI, mg/dL	151 ± 35	148 ± 33	0.557
Apolipoprotein B, mg/dL	100 ± 27	100 ± 38	0.469
Fibrinogen, mg/dL	361 (319–423)	353 (316–399)	0.327
PAI-1, pg/mL	74.5 (55.4–138.8)	81.9 (58.2–137.7)	0.713
IL-6, pg/mL	0.28 (0.08–0.64)	0.54 (0.28–1.4)	0.012
IL-1β, pg/dL	14 (4.0–23.0)	22 (4.6–55.7)	0.003
hsCRP, mg/L	0.54 (0.1–3.1)	0.56 (0.14–3.0)	0.980
TNFα, pg/mL	1.45 (0.34–4.81)	2.9 (1.06–5.84)	0.031

Number of individuals is in parenthesis. Results are shown as mean ± SD or median (interquartile range) and compared by Mann-Whitney Rank sum test. BMR: basal metabolic rate; CAD: coronary artery disease; HDL: high-density lipoprotein; LDL: low-density lipoprotein; hsCRP: high sensitive C reactive protein; IL-6: interleukin 6; IL-1β: interleukin 1beta; PAI-1: plasminogen activator inhibitor-1; sLEPR: soluble leptin receptor; TNFα: tumor necrosis factor alpha; VLDL: very low-density lipoprotein.

In this study, BMI was positively correlated with circulating levels of pro-inflammatory such as fibrinogen, PAI-1, hsCRP, IL-1β, and IL-6 and TNFα. Moreover, altered levels in most of these pro-inflammatory markers were associated with increased risk for obesity. These results confirm previous studies, which suggested that the expansion of the adipose tissue favours the development of a pro-inflammatory, diabetogenic and atherogenic status [3,4].

We also found that variability in TNFα, IL-1β and IL-6 circulating levels was associated with *ADIPOQ* 45 T > G SNP, suggesting a role of this variant in regulation of the pro-inflammatory status in obese subjects. Guzman-

Table 5 Relationship of *IL6* -174G > C with anthropometric, metabolic and inflammatory variables

Variable	Genotypes		P-value
	GG (112)	GC + CC (137)	
Body mass index, kg/m ²	31.2 ± 6.1	30.7 ± 6.3	0.528
Waist circumference, cm	96.4 ± 16.1	92.6 ± 15.7	0.067
Waist- hip ratio	0.87 ± 0.10	0.85 ± 0.10	0.164
Body fat, %	35.5 ± 6.0	36.4 ± 6.2	0.246
BMR, kcal	1644 ± 373	1543 ± 298	0.057
Leptin, ng/mL	21.8 ± 18.6	18.6 ± 15.6	0.394
sLEPR, ng/mL	22.4 ± 23.3	19.2 ± 18.8	0.615
Adiponectin, µg/mL	24.0 ± 24.0	24.5 ± 25.9	0.755
Glucose, mg/dL	103 ± 31	102 ± 21	0.072
HbA1c, %	6.0 ± 1.4	5.9 ± 1.1	0.876
Insulin, mU/L	17.7 ± 13.6	14.3 ± 9.8	0.237
HOMA-β	57.0 ± 40.3	47.4 ± 34.4	0.148
HOMA-IR	4.9 ± 5.3	3.7 ± 2.7	0.379
Total cholesterol, mg/dL	205 ± 42	204 ± 36	0.755
HDL cholesterol, mg/dL	56 ± 19	54 ± 12	0.823
LDL cholesterol, mg/dL	122 ± 34	127 ± 31	0.241
VLDL cholesterol, mg/dL	26 ± 19	25 ± 13	0.759
Triglycerides, mg/dL	132 ± 94	124 ± 64	0.759
Apolipoprotein AI, mg/dL	152 ± 35	148 ± 34	0.753
Apolipoprotein B, mg/dL	101 ± 27	100 ± 34	0.462
Fibrinogen, mg/dL	355 (317–407)	365 (319–420)	0.325
PAI-1, pg/mL	82.7 (57.5–133.5)	75.2 (56.3–145.9)	0.991
IL-6, pg/mL	0.48 (0.08–0.64)	0.28 (0.08–0.96)	0.288
IL-1β, pg/dL	15.0 (4.0–45.0)	13.5 (4.0–32.0)	0.033
hsCRP, mg/L	0.57 (0.13–3.2)	0.52 (0.10–2.94)	0.808
TNFα, pg/mL	2.62 (0.41–4.92)	2.29 (0.49–5.18)	0.666

Number of individuals is in parenthesis. Results are shown as mean ± SD or median (interquartile range) and compared by Mann-Whitney Rank sum test. BMR: basal metabolic rate; CAD: coronary artery disease; HDL: high-density lipoprotein; LDL: low-density lipoprotein; hsCRP: high sensitive C reactive protein; IL-6: interleukin 6; IL-1β: interleukin 1beta; PAI-1: plasminogen activator inhibitor-1; sLEPR: soluble leptin receptor; TNFα: tumor necrosis factor alpha; VLDL: very low-density lipoprotein.

Ornelas et al. have also found an association of *ADIPOQ* 45 T > G SNP with a pattern of fat distribution and correlations with inflammation markers, but not directly related to the genotypes [26].

IL6 -174G > C SNP did not influence the IL-6 circulating levels as it has been shown in a study with Spanish adolescents that demonstrated an association between fat mass and cardiovascular risk factor [37]. Conversely, in Italian Caucasian females, increased plasma levels of IL-6 were found in obese individuals carrying GG genotype suggesting that fat mass is a major determinant of an increase in IL-6 production and insulin resistance [38].

IL-1 β levels were influenced by *IL6* -174G > C SNP in this sample, with high levels found in individuals carrying GG genotype compared to C allele (GC + CC genotypes) carriers. In a similar way, Mendoza-Carrera et al. found an association between C allele and low hsCRP levels in the overweight adolescents [35]. Moreover Ramírez-López et al. reported a relationship between GCG/GCG haplotype from *IL6* -597G > A, -572G > C and -174G > C SNPs and high hsCRP [39]. These results are suggestive of a protective effect of the *IL6* -174G > C variant on inflammatory status of obese individuals.

Even though the results from this study are interesting the lack of association between *ADIPOQ* variant and obesity and adiponectin levels may be influenced by the heterogeneity and size of our sample population and also environmental factors.

Conclusion

The results from this study confirm that obesity is associated with cardiometabolic alterations, leptin resistance and a pro-inflammatory status. Our results are suggestive that *ADIPOQ* and *IL6* polymorphisms contribute to cardiometabolic risk in obese individuals; however, more studies using higher sample sizes are needed to confirm the associations observed in this work.

Abbreviations

ADIPOQ: Adiponectin; BMI: Body mass index; BMR: Basal metabolic rate; CAD: Coronary artery disease; CVD: Cardiovascular disease; hsCRP: High sensitive C reactive protein; IL-1 β : Interleukin 1 beta; IL-6: Interleukin 6; LEPR: Leptin receptor; PAI-1: Plasminogen activator inhibitor-1; sLEPR: Soluble leptin receptor; SNP: Single nucleotide polymorphism; T2DM: Type 2 diabetes mellitus; TNF α : Tumor necrosis factor alpha; WHR: Waist-to-hip ratio.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

All authors have made substantial contributions. RO, MH and RDCH designed the study. ELD and MMSB conducted the clinical evaluations. RO, TIM, CMF and MCS conducted anthropometric measurements, and laboratory and genetic analyses. AC conducted the statistical analyses. RO, AC, MH and RDC reviewed, corrected and helped finalize the manuscript. All authors read and approved the final manuscript.

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References

- Bray GA. Obesity: the disease. *J Med Chem*. 2006;49:4001–7.
- Finucane MM, Stevens GA, Cowan MJ, Danaei G, Lin JK, Paciorek CJ, et al. National, regional, and global trends in body-mass index since 1980: systematic analysis of health examination surveys and epidemiological studies with 960 country-years and 9.1 million participants. *Lancet*. 2011;377:557–67.
- Leal Vde O, Mafra D. Adipokines in obesity. *Clin Chim Acta*. 2013;419:87–94.
- Blüher M. Clinical relevance of adipokines. *Diabetes Metab J*. 2012;36:317–27.
- Yoo HJ, Choi KM. Adipokines as a novel link between obesity and atherosclerosis. *World J Diabetes*. 2014;5:357–63.
- Turer AT, Scherer PE. Adiponectin: mechanistic insights and clinical implications. *Diabetologia*. 2012;55:2319–26.
- Hoene M, Weigert C. The role of interleukin-6 in insulin resistance, body fat distribution and energy balance. *Obes Rev*. 2008;9:20–9.
- Li LL, Kang XL, Ran XJ, Wang Y, Wang CH, Huang L, et al. Associations between 45 T/G polymorphism of the adiponectin gene and plasma adiponectin levels with type 2 diabetes. *Clin Exp Pharmacol Physiol*. 2007;34:1287–90.
- Yu Z, Han S, Cao X, Zhu C, Wang X, Guo X. Genetic polymorphisms in adipokine genes and the risk of obesity: a systematic review and meta-analysis. *Obesity (Silver Spring)*. 2012;30:396–406.
- Sone Y, Yamaguchi K, Fujiwara A, Kido T, Kawahara K, Ishiwaki A, et al. Association of lifestyle factors, polymorphisms in adiponectin, perilipin and hormone sensitive lipase, and clinical markers in Japanese males. *J Nutr Sci Vitaminol (Tokyo)*. 2010;56:123–31.
- Tabatabaei-Malazy O, Hasani-Ranjbar S, Amoli MM, Heshmat R, Sajadi M, Derakhshan R, et al. Gender-specific differences in the association of adiponectin gene polymorphisms with body mass index. *Rev Diabet Stud*. 2010;7:241–6.
- Siitonen N, Pulkkinen L, Lindström J, Kolehmainen M, Eriksson JG, Venojärvi M, et al. Association of ADIPOQ gene variants with body weight, type 2 diabetes and serum adiponectin concentrations: the Finnish Diabetes Prevention Study. *BMC Med Genet*. 2011;12:5.
- Park JW, Park J, Jee SH. ADIPOQ gene variants associated with susceptibility to obesity and low serum adiponectin levels in healthy Koreans. *Epidemiol Health*. 2011;33:e2011003.
- Molerés A, Rendo-Urteaga T, Zulet MA, Marcos A, Campoy C, Garagorri JM, et al. Obesity susceptibility loci on body mass index and weight loss in Spanish adolescents after a lifestyle intervention. *J Pediatr*. 2012;161:466–70.
- Oliveira CS, Saddi-Rosa P, Crispim F, Canani LH, Gerchman F, Giuffrida FM, et al. Association of ADIPOQ variants, total and high molecular weight adiponectin levels with coronary artery disease in diabetic and non-diabetic Brazilian subjects. *J Diabetes Complications*. 2012;26:94–8.
- Cardellini M, Perego L, D'Adamo M, Marini MA, Procopio C, Hribal ML, et al. C-174G polymorphism in the promoter of the interleukin-6 gene is associated with insulin resistance. *Diabetes Care*. 2005;28:2007–12.
- Goyenechea E, Parra D, Martínez JA. Impact of interleukin 6–174G > C polymorphism on obesity-related metabolic disorders in people with excess in body weight. *Metabolism*. 2007;56:1643–8.
- Strandberg L, Mellström D, Ljunggren O, Grundberg E, Karlsson MK, Holmberg AH, et al. IL6 and IL18 polymorphisms are associated with fat mass in older men: the MrOS Study Sweden. *Obesity (Silver Spring)*. 2008;16:710–3.
- Gupta A, Gupta V, Singh AK, Tiwari S, Agrawal S, Natu SM, et al. Interleukin-6 G-174C gene polymorphism and serum resistin levels in North Indian women: potential risk of metabolic syndrome. *Hum Exp Toxicol*. 2011;30:1445–53.
- Popko K, Gorska E, Demkow U. Influence of interleukin-6 and G174C polymorphism in IL-6 gene on obesity and energy balance. *Eur J Med Res*. 2010;15 Suppl 2:123–7.
- Bastien M, Poirier P, Lemieux I, Després JP. Overview of epidemiology and contribution of obesity to cardiovascular disease. *Prog Cardiovasc Dis*. 2014;56:369–81.
- Knight ZA, Hannan KS, Greenberg ML, Friedman JM. Hyperleptinemia is required for the development of leptin resistance. *PLoS One*. 2010;5:e11376.
- Martin SS, Qasim A, Reilly MP. Leptin resistance: a possible interface of inflammation and metabolism in obesity-related cardiovascular disease. *J Am Coll Cardiol*. 2008;52:1201–10.
- Kahn SE, Hull RL, Utzschneider KM. Mechanisms linking obesity to insulin resistance and type 2 diabetes. *Nature*. 2006;444:840–6.
- Mousavinasab F, Tähtinen T, Jokelainen J, Koskela P, Vanhala M, Oikarinen J, et al. Common polymorphisms (single-nucleotide polymorphisms SNP + 45

- and SNP + 276) of the adiponectin gene regulate serum adiponectin concentrations and blood pressure in young Finnish men. *Mol Genet Metab.* 2006;87:147–51.
26. Guzman-Ornelas MO, Chavarria-Avila E, Munoz-Valle JF, Armas-Ramos LE, Castro-Albarran J, Aguilar Aldrete ME, et al. Association of ADIPOQ +45 T > G polymorphism with body fat mass and blood levels of soluble adiponectin and inflammation markers in a Mexican-Mestizo population. *Diabetes Metab Syndr Obes.* 2012;5:369–78.
 27. Al-Daghri NM, Al-Attas OS, Alokail MS, Alkharfy KM, Hussain T, Yakout S, et al. Adiponectin gene polymorphisms (T45G and G276T), adiponectin levels and risk for metabolic diseases in an Arab population. *Gene.* 2012;493:142–7.
 28. Wu J, Liu Z, Meng K, Zhang L. Association of adiponectin gene (ADIPOQ) rs2241766 polymorphism with obesity in adults: a meta-analysis. *PLoS One.* 2014;9:e95270.
 29. Cohen SS, Gammon MD, North KE, Millikan RC, Lange EM, Williams SM, et al. ADIPOQ, ADIPOR1, and ADIPOR2 polymorphisms in relation to serum adiponectin levels and BMI in black and white women. *Obesity (Silver Spring).* 2011;19:2053–62.
 30. Rizk NM, El-Menyar A, Marei I, Sameer M, Musad T, Younis D, et al. Association of adiponectin gene polymorphism (+T45G) with acute coronary syndrome and circulating adiponectin levels. *Angiology.* 2013;64:257–65.
 31. Wassel CL, Pankow JS, Jacobs Jr DR, Steffes MW, Li N, Schreiner PJ. Variants in the adiponectin gene and serum adiponectin: the Coronary Artery Development in Young Adults (CARDIA) Study. *Obesity (Silver Spring).* 2010;18:2333–8.
 32. Antonopoulos AS, Tousoulis D, Antoniadis C, Miliou A, Hatzis G, Papageorgiou N, et al. Genetic variability on adiponectin gene affects myocardial infarction risk: the role of endothelial dysfunction. *Int J Cardiol.* 2013;168:326–30.
 33. Qi L, Zhang C, van Dam RM, Hu FB. Interleukin-6 genetic variability and adiposity: associations in two prospective cohorts and systematic review in 26,944 individuals. *J Clin Endocrinol Metab.* 2007;92:3618–5.
 34. Pyrzak B, Wisniewska A, Majcher A, Popko K, Wasik M, Demkow U. Association between metabolic disturbances and G-174C polymorphism of interleukin-6 gene in obese children. *Eur J Med Res.* 2009;14 Suppl 4:196–200.
 35. Mendoza-Carrera F, Ramírez-López G, Ayala-Martínez NA, García-Zapién AG, Flores-Martínez SE, Sánchez-Corona J. Influence of CRP, IL6, and TNFA gene polymorphisms on circulating levels of C-reactive protein in Mexican adolescents. *Arch Med Res.* 2010;41:472–7.
 36. Bouhaha R, Baroudi T, Ennafaa H, Vaillant E, Abid H, Sassi R, et al. Study of TNFalpha -308G/A and IL6-174G/C polymorphisms in type 2 diabetes and obesity risk in the Tunisian population. *Clin Biochem.* 2010;43:549–52.
 37. Molerés A, Rendo-Urteaga T, Azcona C, Martínez JA, Gómez-Martínez S, Ruiz JR, et al. IL6 gene promoter polymorphism (-174G/C) influences the association between fat mass and cardiovascular risk factors. *J Physiol Biochem.* 2009;65:405–13.
 38. Di Renzo L, Bertoli A, Bigioni M, Del Gobbo V, Premrov MG, Calabrese V, et al. Body composition and -174G/C interleukin-6 promoter gene polymorphism: association with progression of insulin resistance in normal weight obese syndrome. *Curr Pharm Des.* 2008;14:2699–706.
 39. Ramírez-López G, Portilla-de Buen E, Sánchez-Corona J, Salmerón-Castro J, Mendoza-Carrera F. Interleukin-6 polymorphisms are associated with obesity and hyperglycemia in Mexican adolescents. *Arch Med Res.* 2013;44:62–8.

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