

RESEARCH

Open Access



Association between serum PCSK9 and coronary heart disease in patients with type 2 diabetes mellitus

Juan Huang^{1†}, Jun-Xu Gu^{2†}, Kun Wang^{3†}, Ai-Min Zhang^{2†}, Ting-Ting Hong², Shan-Shan Li², Xiao-Qin Yao¹, Ming Yang⁴, Yue Yin², Na Zhang², Ming Su², Jia-Jia Hu^{2*}, Xue-Zhi Zhang^{1*} and Mei Jia^{2*} 

Abstract

Background and aims Proprotein convertase subtilisin/kexin type 9 (PCSK9) is considered a new biomarker for atherosclerosis, but its ability to predict cardiovascular outcomes has been controversial. This study aimed to address the lack of data on PCSK9, coronary heart disease (CHD) severity, and major cardiovascular events (MACEs) in patients with type 2 diabetes mellitus (T2DM).

Methods A total of 2984 T2DM patients underwent selective coronary angiography, and their serum PCSK9 levels were measured using enzyme-linked immunosorbent assay. Correlation and logistic regression analyses were performed to investigate the association between PCSK9 expression and CHD severity. This study used Cox regression analysis to assess the association between circulating PCSK9 levels and the risk of MACEs.

Results Circulating PCSK9 levels were significantly higher in the CHD group than in the non-CHD group [554.62 (265.11) ng/mL vs. 496.86 (129.05) ng/mL, $p < 0.001$]. Circulating PCSK9 levels positively correlated with CHD severity (diseased vessels: $r = 0.35$, $p < 0.001$; Gensini score: $r = 0.46$, $p < 0.001$). Elevated PCSK9 levels are an independent risk factor for CHD risk and severity (CHD group vs. non-CHD group: OR = 2.829, 95% CI: 1.771–4.520, $p < 0.001$; three vessel disease group vs. one vessel disease group: OR = 4.800, 95% CI: 2.387–9.652, $p < 0.001$; high GS group vs. low GS group: OR = 5.534, 95% CI: 2.733–11.208, $p < 0.001$). Through a six-year follow-up and multivariate Cox regression analysis, elevated circulating PCSK9 levels were found to be independently associated with MACEs in all participants (HR: 3.416, 95% CI: 2.485–4.697, $p < 0.001$; adjusted HR: 2.780, 95% CI: 1.930–4.004, $p < 0.001$).

Conclusions Serum PCSK9 levels were positively correlated with multi-vessel CHD and Gensini score. Elevated circulating PCSK9 levels are an independent risk factor for CHD and increased incidence of MACEs in T2DM.

[†]Juan Huang, Jun-Xu Gu, Kun Wang and Ai-Min Zhang contributed equally to this work and share first authorship.

*Correspondence:

Jia-Jia Hu
18895660269@163.com
Xue-Zhi Zhang
zhangxuezhi@pkuih.edu.cn
Mei Jia
jiamei00718@sina.com

Full list of author information is available at the end of the article



© The Author(s) 2023. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

Keywords PCSK9, T2DM, Coronary heart disease, Cardiovascular events

Introduction

Coronary heart disease (CHD) is a prevalent cardiovascular disease and is the leading cause of disability and death in humans [1–3]. Over the past half a century, several studies have shown that diabetes, hypertension, dyslipidemia, and smoking are possible risk factors for cardiovascular disease that can be used for the early assessment of the risk for cardiovascular disease [4, 5]. Type 2 diabetes mellitus (T2DM) has a variety of causes that lead to insufficient insulin secretion or the inability of the body to effectively use insulin, resulting in a sustained increase in blood sugar levels and impaired carbohydrate, lipid, and protein metabolism. It mainly occurs in adults with obesity, and the number of females with T2DM is greater than that of males with T2DM [6, 7]. T2DM patients are prone to lipid metabolism disorders, thereby having increased risk of cardiovascular diseases [8, 9].

The proprotein convertase subtilisin/kexin type 9 (PCSK9) is a member of the proprotein convertase enzyme family [10, 11]. PCSK9 is a serine protease mainly derived from the liver and intestines; however, according to animal studies, the primary source of circulating PCSK9 is the liver [12, 13]. PCSK9 regulates the degradation of low-density lipoprotein receptors (LDLR) on the surface of liver cells via the endosome/lysosome pathway, revealing a new concern in low-density cholesterol homeostasis. In addition to this classical pathway, recent research has also found that PCSK9 plays other roles in the development of atherosclerosis via non-classical mechanisms, such as inflammation, apoptosis and immune pathways [11, 14, 15].

In recent years, many studies have shown that PCSK9 can promote the occurrence and development of vascular atherosclerosis by promoting an inflammatory response and endothelial dysfunction and inhibiting platelet activation [16, 17]. Several large-scale clinical trials have found that human circulating PCSK9 monoclonal antibodies can significantly reduce serum low-density lipoprotein cholesterol (LDL-C) levels and demonstrate outstanding potential for reducing future cardiovascular events. Most importantly, circulating PCSK9 has been recognized as an adverse cardiovascular risk factor in patients with CAD [18, 19]. Owing to the close relationship between lipid and glucose metabolism, an increasing number of studies have found a close relationship between PCSK9 and diabetes [20, 21]. However, the pathogenicity of PCSK9 in T2DM patients and T2DM patients with CHD has not been completely established.

Therefore, this study aims to investigate serum PCSK9 levels and severity of CHD in stable CHD patients with

T2DM, as well as the risk of long-term cardiovascular events.

Methods

Study population

This is a cohort study. This study enrolled 2984 T2DM patients (1732 males, 1252 females) from Peking University International Hospital and Peking University People's Hospital between October 2013 and September 2016. The participants included 1776 patients (1046 males, 730 females) with CHD and 1208 (686 males, 522 females) without CHD. The follow-up procedures were performed every six months by experienced nurses or doctors through telephone or face-to-face interviews. This is illustrated in the flowchart (Fig. 1). Major cardiovascular events (MACEs) include cardiovascular mortality, non-fatal myocardial infarction (MI), non-fatal stroke, heart failure, and hospitalization for unstable angina. The longest follow-up time among all patients was six years.

All patients diagnosed with T2DM were selected according to the following criteria set by the American Diabetes Association: (1) currently receiving treatment of oral hypoglycemic medicine or insulin, (2) self-reporting to the clinician that they have a history of type 2 diabetes, (3) repeated fasting plasma glucose (FPG) greater than 7.0 mmol/L, or (4) glycated hemoglobin A1c (HbA1c) $\geq 6.5\%$.

The diagnostic criteria for patients with CHD were based on the results of coronary angiography (CAG) performed by Peking University International Hospital or Peking University People's Hospital, which is defined as at least one major coronary artery occlusion or stenosis exceeding 50%, and the severity of CHD was evaluated using the Gensini score (GS).

Exclusion criteria included: (1) a history of coronary artery bypass surgery; (2) percutaneous coronary intervention within three months; (3) acute coronary syndrome within six months; (4) cardiomyopathy, valvular heart disease, or chronic heart failure; (5) pulmonary heart disease; (6) severe liver and kidney dysfunction; or (7) any known inflammatory or infectious disease, or confirmed, or suspected cancer.

This study complied with the Declaration of Helsinki and was approved by the Peking University People's Hospital Research Ethics Committee (No.2018PHB155-01). Written informed consent was obtained from all the enrolled patients.

Bioinformatics analysis of PCSK9

Protein and protein network analysis STRING (<https://string-preview.org/>) was used to construct and analyze

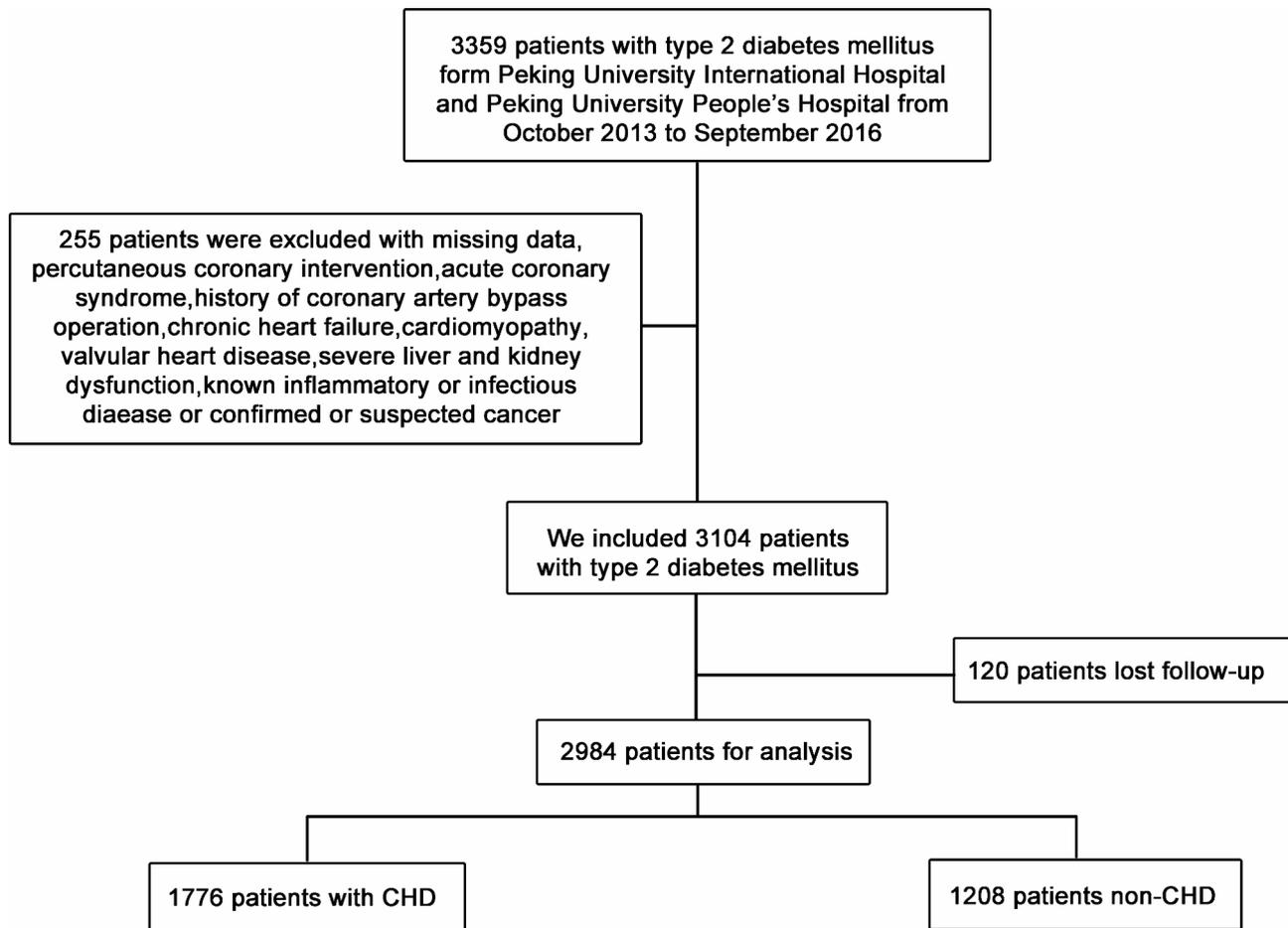


Fig. 1 The flow chart of the patients' selection process

protein and protein networks based on PCSK9. Enriched Gene Ontology and Kyoto Encyclopedia of Genes and Genomes pathways were evaluated using gene set enrichment analysis.

Conventional clinical and laboratory indicator tests

Blood samples were collected in the morning after fasting for 12 h. Serum samples were prepared by centrifugation at 3500 rpm for 10 min at 10–15 °C and were stored at –80 °C.

The circulating levels of PCSK9 were measured using the ELISA kit (Duduo Biotechnology Co., Ltd. China), according to the manufacturer's instruction. Fasting blood glucose (FBG), hypersensitive C-reactive protein (hs-CRP), homocysteine (HCY), and serum lipid profiles, including triglycerides (TG), total cholesterol (TC), LDL-C, and high-density lipoprotein cholesterol (HDL-C), were analyzed using the Beckman AU5832 analyzer (Beckman Coulter Inc., USA). Lipoprotein (a) [Lp(a)] was measured using the latex enhanced immunoturbidimetry Lp(a) kit (Roche Inc., Germany). Apolipoprotein A-1 (apoA1) and B (apoB) were measured

using immunoturbidimetry (Daiichi Pure Chemicals Co., Ltd., Tokyo). Small dense low-density lipoprotein cholesterol (sdLDL-C) was detected using sdLDL-C direct quantitative analysis kits (Denka Seiken Co., Ltd. Japan). Hemoglobin A1c (HbA_{1c}) levels were determined using high-performance liquid chromatography (Trinity Biotech Inc., USA).

Statistical analyses

The one-sample Kolmogorov Smirnov test was used to analyze the distribution of all quantitative variables. Normally distributed data are presented as mean ± standard deviation, and the Student's t-test and the analysis of variance were used to compare the differences between groups. Continuous data with non-normal distribution are reported as medians (inter-quartile ranges), and differences between various groups were compared using the Mann–Whitney U test and Kruskal–Wallis test. Categorical data were presented as percentages (%) and compared using the chi-square test. Spearman's correlation analysis was used to calculate the correlation coefficient between

the circulating PCSK9 levels and CHD severity. The relationship between PCSK9 and CHD severity was analyzed using univariate and multivariate logistic analyses. The Kaplan–Meier method was used to estimate the survival rate of MACEs in the groups. Hazard ratios (HR) and 95% confidence intervals (CI) for the relationship between circulating PCSK9 levels and the incidence of MACEs were generated using univariate and multivariate Cox regression analyses. $p < 0.05$ was considered statistically significant. SPSS 22.0 for Windows (SPSS Inc., USA) and GraphPad Prism 7 (GraphPad Software Inc., USA) were used for statistical analyses.

Results

Baseline characteristics of all patients

We initially analyzed the baseline characteristics of all participants. Table 1 presents the demographic, anthropometric, and biochemical characteristics of the study population. There were no significant differences between the CHD and non-CHD groups in terms of sex (male), age, body mass index (BMI), percentage of hypertension, alcohol consumption,

smoking, family history of diabetes mellitus (MD), FBG levels, HbA_{1c}, TG, hs-CRP and HCY. In contrast, the CHD group showed a higher percentage of a family history of CHD than the non-CHD group. In addition, a statistically significant increase in ApoB, TC, LDL-C, Lp(a), sdLDL-C, and PCSK9 levels [554.62 (265.11) ng/ml vs. 496.86 (129.05) ng/ml, $p < 0.001$] was observed, whereas ApoA1 and HDL-C levels decreased in the CHD group.

Next, we divided all the patients into four subgroups based on the quartile of PCSK9 levels (Table 2). ApoA1 and HDL-C levels were higher in the lowest quartile than in the highest quartile of PCSK9 levels (all $p < 0.05$). Furthermore, the levels of Lp(a), TC, LDL-C, ApoB, HCY, and sdLDL-C were significantly higher in participants in the highest quartile of PCSK9 levels than in those in the lowest quartile of PCSK9 levels (all $p < 0.05$). In addition, there were no significant differences in age, sex (male), BMI, percentage of hypertension, smoking, alcohol consumption, family history of CHD, family history of DM, FBG levels, HbA_{1c}, TG and hs-CRP between the two groups (all $p > 0.05$).

Table 1 Baseline characteristics in type 2 diabetic patients

	Total	CHD group	Non-CHD group	<i>p</i> value
Clinical characteristics				
N (%)	2984	1776 (59.52%)	1208 (40.48%)	–
Age (years)	57.48 ± 9.93	57.81 ± 9.74	56.99 ± 10.19	0.222
Male (%)	1732 (58.04%)	1046 (58.90%)	686 (56.78%)	0.252
BMI (kg/m ²)	25.23 ± 3.07	25.06 ± 2.90	25.48 ± 3.31	0.131
Hypertension (%)	2084 (69.53%)	1252 (70.50%)	832 (68.87%)	0.344
Smoking (%)	838 (28.08%)	481 (27.08%)	357 (29.55%)	0.141
Alcohol consumption (%)	887 (29.73%)	541 (30.46%)	346 (28.64%)	0.286
Family history of CHD (%)	851 (28.52%)	531 (29.90%)	320 (26.49%)	0.043
Family history of MD (%)	1135 (38.04%)	655 (36.88%)	480 (39.73%)	0.115
Laboratory variables				
FBG (mmol/L)	7.69 ± 1.16	7.73 ± 1.21	7.64 ± 1.08	0.219
HbA _{1c} (%)	7.52 ± 1.10	7.55 ± 1.15	7.48 ± 1.02	0.367
ApoB (mg/dL)	85.10 (26.85)	87.00 (36.60)	84.16 (20.16)	0.004
ApoA1 (mg/dL)	144.1 (37.55)	142.50 (41.50)	145.65 (31.58)	0.014
Total cholesterol (mmol/L)	4.44 (1.34)	4.46 (1.72)	4.41 (0.98)	0.009
Triglycerides (mmol/L)	1.32 (0.88)	1.32 (0.82)	1.32 (0.94)	0.395
HDL-C (mmol/L)	1.14 (0.35)	1.13 (0.30)	1.15 (0.39)	0.005
LDL-C (mmol/L)	2.73 (0.94)	2.79 (1.37)	2.71 (0.67)	0.011
Lp(a) (nmol/L)	39.24 (39.77)	40.51 (48.21)	36.99 (33.45)	0.001
hs-CRP (mg/L)	1.38 (1.69)	1.57 (1.80)	1.23 (1.56)	0.386
HCY (umol/L)	11.65 (7.69)	11.77 (8.51)	11.45 (6.60)	0.091
sdLDL-C (mmol/L)	0.74 (0.37)	0.76 (0.47)	0.71 (0.31)	0.005
PCSK9 (ng/mL)	521.98 (209.78)	554.62 (265.11)	496.86 (129.05)	< 0.001

Data are reported as means ± SD or n(%), median (interquartile ranges). SD: Standard deviation

BMI: body mass index; FPG: fasting plasma glucose; HbA_{1c}: Hemoglobin A_{1c}; apoB: apolipoprotein B; apoA1: apolipoprotein A1; HDL-C: high density lipoprotein cholesterol; LDL-C: low density lipoprotein cholesterol; Lp(a): lipoprotein (a); Hs-CRP: hypersensitive C-reactive protein; HCY: homocysteine; sdLDL-C: small dense low-density lipoprotein cholesterol; PCSK9: proprotein convertase enzyme subtilisin/kexin type 9

Statistical analysis was performed with the Student's t test or Mann-Whitney U test and with Chi-square test for categorical variables

Table 2 Baseline characteristics of patients with type 2 diabetes at different circulating PCSK9 levels

Variables	PCSK9 concentration (ng/mL)				p value
	Q1: < 432.98	Q2: 432.98–521.98	Q3: 521.98–621.24	Q4: > 621.24	
Clinical characteristics					
N (%)	748 (25.07%)	744 (24.93%)	746 (25.00%)	746 (25.00%)	–
Age (years)	57.07 ± 9.19	57.59 ± 9.94	56.94 ± 10.03	58.31 ± 10.33	0.440
Male (%)	437 (58.42%)	419 (56.32%)	451 (60.46%)	425 (56.97%)	0.378
BMI (kg/m ²)	24.91 ± 2.82	25.55 ± 3.54	25.47 ± 2.98	24.97 ± 2.85	0.203
Hypertension (%)	501 (66.98%)	532 (71.51%)	541 (72.52%)	510 (68.36%)	0.066
Smoking (%)	220(29.41%)	193(25.94%)	215(28.82%)	210(28.15%)	0.467
Alcohol consumption (%)	232(31.02%)	236(31.72%)	216(28.95%)	203(27.21%)	0.214
Family history of CHD (%)	194(25.94%)	201(27.02%)	220(29.49%)	236(31.64%)	0.067
Family history of MD (%)	272(36.36%)	293(39.38%)	280(37.53%)	290(38.87%)	0.623
Laboratory variables					
FPG (mmol/L)	7.62 ± 1.10	7.59 ± 1.11	7.80 ± 1.15	7.78 ± 1.21	0.057
HbA1c (%)	7.44 ± 1.06	7.44 ± 1.02	7.58 ± 1.10	7.62 ± 1.19	0.181
ApoB (mg/dL)	83.10 (26.80)	81.56 (23.96)	84.48 (25.47)	92.95 (31.95) ^{abc}	<0.001
ApoA1 (mg/dL)	150.70 (33.05)	145.71 (48.11)	142.05 (32.93) ^a	140.50 (32.99) ^{ab}	0.001
Total cholesterol (mmol/L)	4.34 (1.42)	4.32 (1.29)	4.41 (1.19)	4.64 (1.49) ^{abc}	<0.001
Triglycerides (mmol/L)	1.33 (0.82)	1.28 (0.96)	1.31 (0.95)	1.36 (0.82)	0.997
HDL-C (mmol/L)	1.16 (0.41)	1.13 (0.38)	1.12 (0.27) ^a	1.12 (0.35) ^a	0.023
LDL-C (mmol/L)	2.66 (1.00)	2.69 (0.81)	2.73 (0.85) ^a	3.02 (1.17) ^{abc}	<0.001
Lp(a) (nmol/L)	30.67 (19.41)	36.54 (26.13) ^a	40.33 (35.36) ^a	76.71 (84.78) ^{abc}	<0.001
hs-CRP (mg/L)	1.33 (1.62)	1.51 (1.85)	1.31 (1.79)	1.34 (1.55)	0.948
HCY (umol/L)	11.21 (6.62)	11.40 (7.14)	11.33 (6.62)	13.56 (9.80) ^{abc}	<0.001
sdLDL-C (mmol/L)	0.73 (0.43)	0.69 (0.43)	0.75 (0.35)	0.79 (0.47) ^{ab}	0.008
PCSK9 (ng/mL)	379.84 (83.46)	480.04 (47.30)	566.52 (44.78)	765.66 (200.58)	

Data are reported as means ± SD or n(%), median (interquartile ranges). SD: Standard deviation

BMI: body mass index; FPG: fasting plasma glucose; HbA1c: Hemoglobin A1c; apoB: apolipoprotein B; apoA1: apolipoprotein A1; HDL-C: high density lipoprotein cholesterol; LDL-C: low density lipoprotein cholesterol; Lp(a): lipoprotein (a); Hs-CRP: hypersensitive C-reactive protein; HCY: homocysteine; sdLDL-C: small dense low-density lipoprotein cholesterol; PCSK9: proprotein convertase enzyme subtilisin/kexin type 9

Statistical analysis was performed with the ANOVA or Kruskal – Wall test and with Chi-square test for categorical variables

a: Shows that the $p < 0.05$ compared with the Q1 group

b: Shows that the $p < 0.05$ compared with the Q2 group

c: Shows that the $p < 0.05$ compared with the Q3 group

Association between PCSK9 and the severity of CHD

To explore the relationship between PCSK9 and CHD severity, we divided the patients in the CHD group into four subgroups based on the quartile of PCSK9 levels (Supplementary Table 1). We observed that the percentage of patients with three-vessel disease (17.30% vs. 31.01% vs. 50.45% vs. 66.52%, $p < 0.001$) and GS [23 (21) vs. 30 (21) vs. 36 (24) vs. 46 (20), $p < 0.001$] increased in the high PCSK9 group, whereas the percentage of patients with one-vessel disease decreased ($p < 0.001$), indicating that PCSK9 levels may be associated with CHD severity. Furthermore, Spearman’s correlation analysis showed that serum PCSK9 levels were significantly and positively associated with diseased vessels and GS ($r = 0.35$, $p < 0.001$; $r = 0.46$, $p < 0.001$, respectively) (Fig. 2).

We also performed a protein–protein interaction analysis on PCSK9 by using STRING and the results were as shown in Supplementary Materials Fig. 1. In addition, GO annotations and KEGG signaling

pathway analysis for these related genes and proteins were provided in Supplementary Materials Fig. 2.

Serum PCSK9 is associated with the risk of CHD in diabetic patients

Based on the CAG examination reports of each patient, diabetic patients with CHD were further divided into one-, two-, and three-vessel disease groups, or low- and high-GS groups, to further evaluate the relationship between PCSK9 and the severity of CHD.

Multivariate binary logistic regression was employed to analyze the relationship between PCSK9 and CHD risk and severity. All participants were divided into subgroups according to PCSK9 level quartiles, and the lowest PCSK9 quartile group was used as a reference to assess the risk and severity of CHD in individuals with different PCSK9 levels.

Univariate logistic regression analysis showed that the ORs for CHD risk and severity were positively associated with PCSK9 levels (CHD group vs. non-CHD group:

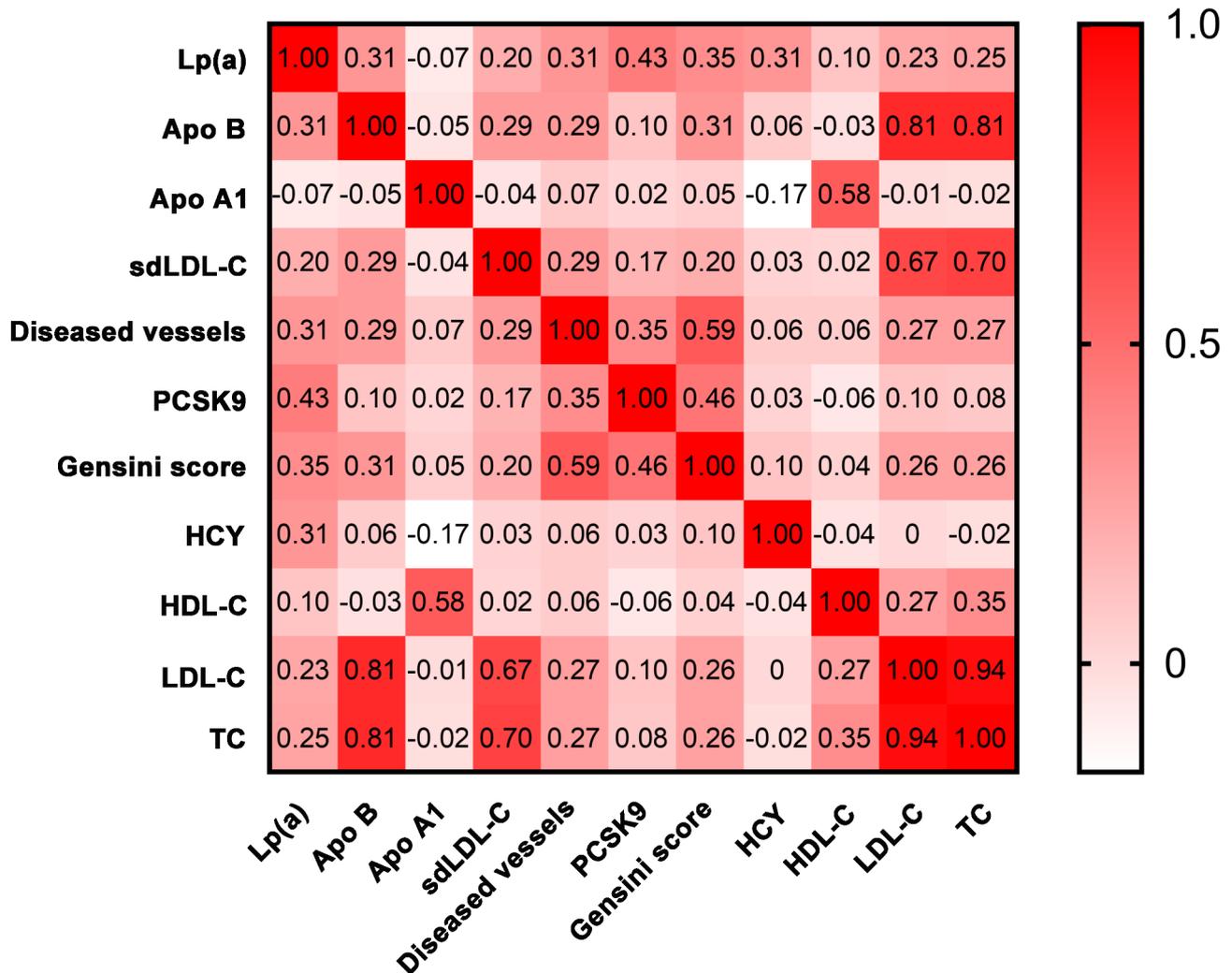


Fig. 2 Correlation between PCSK9 and severity of coronary heart disease (diseased vessels: $r=0.35, p<0.001$; Gensini score: $r=0.46, p<0.001$)

OR=3.308, 95% CI: 2.137–5.121, $p<0.001$; three vessel disease group vs. one vessel disease group: OR=6.290, 95% CI: 3.349–11.814, $p<0.001$; high GS group vs. low GS group: OR=5.889, 95% CI: 3.504–9.899, $p<0.001$) (Fig. 3, A, C, E). Furthermore, the upward trend remained even after adjusting for age, sex, BMI, hypertension, smoking, alcohol consumption, family history of CHD, Family history of DM, FPG, HbA_{1c}, ApoB, ApoA1, TC, TG, HDL-C, LDL-C, Lp(a), hs-CRP, HCY and sdLDL-C levels compared with those in the first quartile of PCSK9 (CHD group vs. non-CHD group: OR=2.829, 95% CI: 1.771–4.520, $p<0.001$; three vessel disease group vs. one vessel disease group: OR=4.800, 95% CI: 2.387–9.652, $p<0.001$; high GS group vs. low GS group: OR=5.534, 95% CI: 2.733–11.208, $p<0.001$) (Fig. 3, B, D, F). Taken together, these data indicate that PCSK9 levels are positively correlated with CHD severity.

PCSK9 and cardiovascular outcomes

After six years of follow-up, 373 (12.50%) individuals developed MACEs. In the non-CHD group, 138 individuals were diagnosed with CHD, and 96 (7.95%) of these patients had MACEs. In the CHD group, 277 (15.60%) patients experienced MACEs. Patients with MACEs had presented with clinical features, such as high percentage of family history of CHD, ApoB, TC, sdLDL-C, and PCSK9 [644.66 (310.41) ng/ml vs. 511.01 (169.12) ng/ml, $p<0.001$] (Table 3). Based on the quartiles of PCSK9 levels, all 2984 individuals were divided into four subgroups to investigate the relationship between the levels of PCSK9 and MACEs risk. As shown in Supplementary Tables 2, 3, 4, 5 and 6, the total incidence of MACEs in the highest PCSK9 quartile group was significantly higher than that in the other three groups, including the non-CHD group (2.90% vs. 8.11% vs. 9.52% vs. 16.98%, $p<0.001$), the CHD group (9.36% vs. 13.54% vs. 14.88% vs. 21.25%, $p<0.001$), all the participants (6.68% vs.

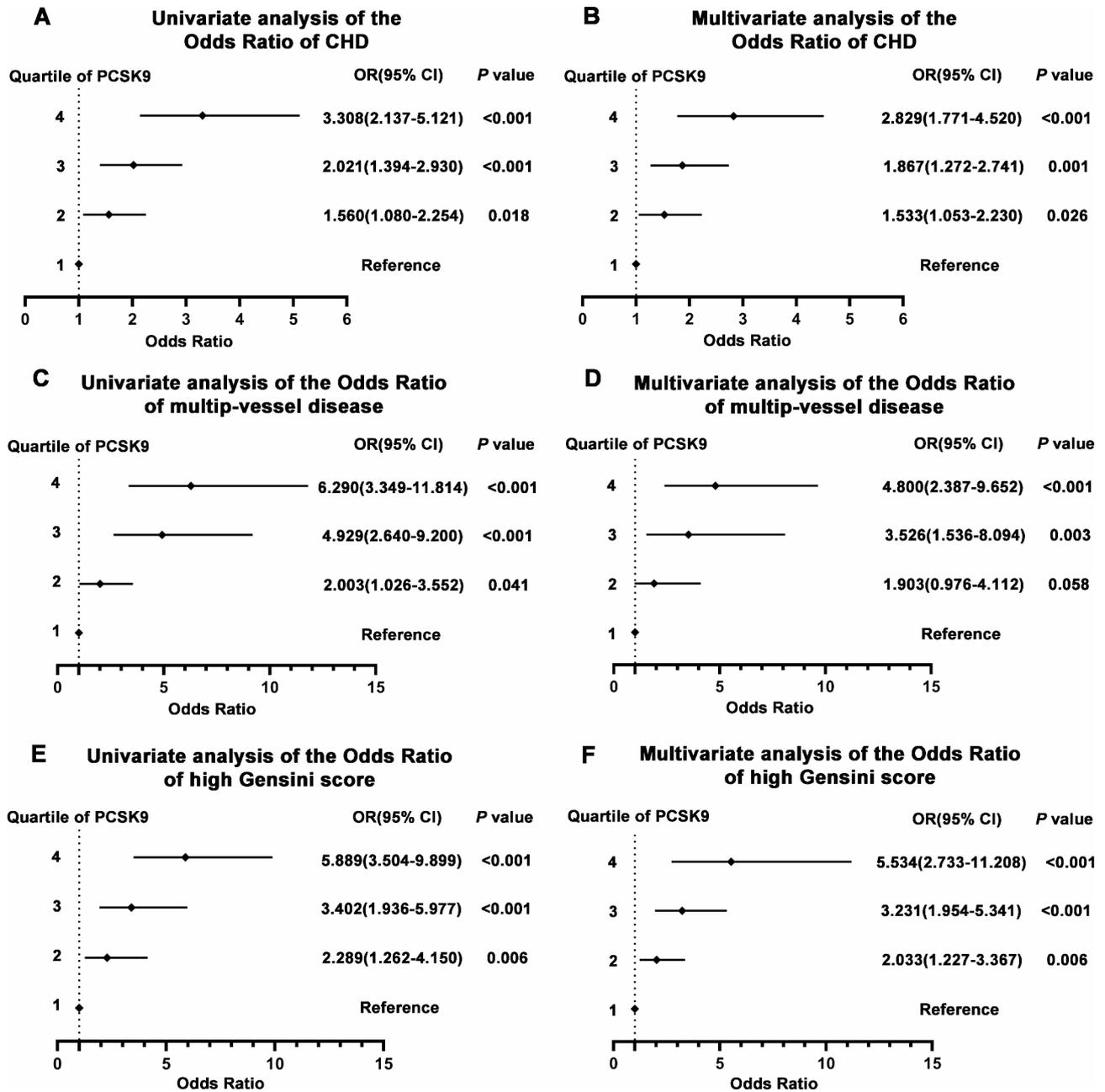


Fig. 3 Logistic regression analysis between circulating PCSK9 quartile and CHD, multiple-vessel disease and high Gensini score. (A) univariate logistic regression analysis between circulating PCSK9 and CHD; (B) multivariate logistic regression analysis between circulating PCSK9 and CHD; (C) univariate logistic regression analysis between circulating PCSK9 and multiple-vessel disease; (D) multivariate logistic regression analysis between circulating PCSK9 and multiple-vessel disease; (E) univariate logistic regression analysis between circulating PCSK9 and Gensini score; (F) multivariate logistic regression analysis between circulating PCSK9 and Gensini score

10.22% vs. 12.47% vs. 20.64%, $p < 0.001$), male participants (6.76% vs. 11.58% vs. 12.91% vs. 20.68%, $p < 0.001$), and female participants (6.58% vs. 8.58% vs. 11.76% vs. 20.59%, $p < 0.001$). Furthermore, Kaplan–Meier analysis showed that the incidence of MACEs in the highest PCSK9 quartile group was higher than that in the other three groups (Fig. 4C, $p < 0.001$). The results were similar

between the non-CHD and CHD groups (Fig. 4A and B, all $p < 0.001$).

According to the univariate Cox regression analysis, there was a 6.070-fold higher risk of MACEs in the highest PCSK9 quartile group than that in the lowest (reference group) in the non-CHD group (HR: 6.070, 95% CI: 2.727–13.512, $p < 0.001$) (Fig. 5A). A significant association was observed after adjusting for confounding factors

Table 3 Baseline characteristics in all subjects with or without MACEs

	Total	MACEs group	Non- MACEs group	p value
Clinical characteristics				
N (%)	2984	373 (12.50%)	2611 (87.50%)	–
Age (years)	57.48 ± 9.93	58.42 ± 11.37	57.34 ± 9.69	0.265
Male (%)	1732 (58.04%)	225 (60.32%)	1507 (57.72%)	0.340
BMI (kg/m ²)	25.23 ± 3.07	24.89 ± 2.62	25.27 ± 3.14	0.329
Hypertension (%)	2084 (69.53%)	276 (73.99%)	1808 (69.25%)	0.062
Smoking (%)	838 (28.08%)	108 (28.95%)	730 (27.96%)	0.689
Alcohol consumption (%)	887 (29.73%)	110 (29.49%)	777 (29.75%)	0.916
Family history of CHD (%)	851 (28.52%)	131 (35.12%)	720 (27.58%)	0.003
Family history of MD (%)	1135 (38.04%)	146 (39.14%)	989 (37.88%)	0.638
Laboratory variables				
FPG (mmol/L)	7.69 ± 1.16	7.73 ± 1.13	7.68 ± 1.15	0.679
HbA1c (%)	7.52 ± 1.10	7.58 ± 1.13	7.51 ± 1.09	0.492
ApoB (mg/dL)	85.10 (26.85)	90.00 (33.90)	84.52 (25.79)	0.008
ApoA1 (mg/dL)	144.1 (37.55)	144.35 (34.57)	144.10 (37.70)	0.874
Total cholesterol (mmol/L)	4.44 (1.34)	4.59 (1.30)	4.43 (1.33)	0.013
Triglycerides (mmol/L)	1.32 (0.88)	1.37 (0.91)	1.31 (0.87)	0.411
HDL-C (mmol/L)	1.14 (0.35)	1.09 (0.37)	1.14 (0.35)	0.257
LDL-C (mmol/L)	2.73 (0.94)	2.73 (1.16)	2.73 (0.92)	0.161
Lp(a) (nmol/L)	39.24 (39.77)	39.51 (41.25)	39.31 (40.03)	0.527
hs-CRP (mg/L)	1.38 (1.69)	1.32 (1.73)	1.39 (1.66)	0.312
HCY (umol/L)	11.65 (7.69)	11.79 (6.88)	11.62 (7.86)	0.941
sdLDL-C (mmol/L)	0.74 (0.37)	0.80 (0.39)	0.73 (0.37)	0.048
PCSK9 (ng/mL)	521.98 (209.78)	644.66 (310.41)	511.01 (169.12)	< 0.001

Data are reported as means ± SD or n(%), median (interquartile ranges). SD: Standard deviation

BMI: body mass index; FPG: fasting plasma glucose; HbA1c: Hemoglobin A1c; apoB: apolipoprotein B; apoA1: apolipoprotein A1; HDL-C: high density lipoprotein cholesterol; LDL-C: low density lipoprotein cholesterol; Lp(a): lipoprotein (a); Hs-CRP: hypersensitive C-reactive protein; HCY: homocysteine; sdLDL-C : small dense low-density lipoprotein cholesterol

Statistical analysis was performed with the Student's t test or Mann-Whitney U test and with Chi-square test for categorical variables

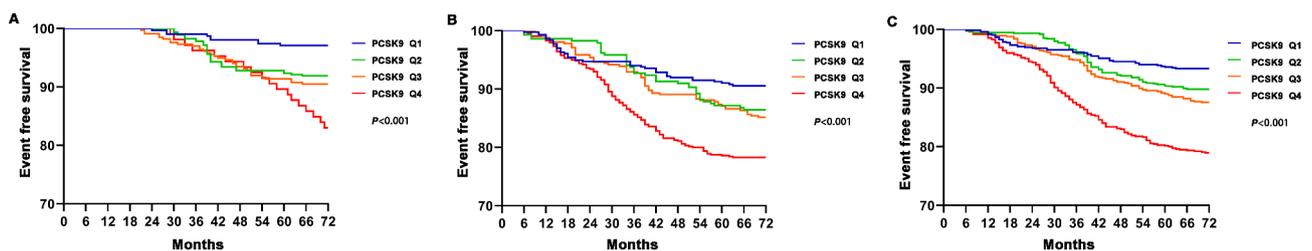


Fig. 4 Kaplan-Meier curve based on circulating PCSK9 quartile values among different patients. (A) in the non-CHD group; (B) in the CHD group; (C) in all participants

(HR: 5.261, 95% CI: 2.226–12.437, $p < 0.001$) (Fig. 5B). In the CHD group and all the participants, the results were consistent with those of the non-CHD group in both univariate and multivariate Cox regression analyses (CHD: HR: 2.464, 95% CI: 1.739–3.490, $p < 0.001$; adjusted HR: 2.238, 95% CI: 1.696–3.420, $p < 0.001$. All subjects: HR: 3.416, 95% CI: 2.485–4.697, $p < 0.001$; adjusted HR: 2.780, 95% CI: 1.930–4.004, $p < 0.001$) (Fig. 5C–F; Supplementary Table 7). These data, indicate that the PCSK9 level is related to MACEs in all the participants.

Discussion

In this study, we investigated the association between circulating PCSK9 and the risk and severity of CHD, by evaluating circulating PCSK9 as a predictive biomarker for cardiovascular events in T2DM patients. We found that circulating PCSK9 was positively associated with the risk of CHD in patients with T2DM and was positively associated with the severity of CHD. We also found that higher PCSK9 levels indicated higher rates of cardiovascular events.

To the best of our knowledge, this is the first large-scale longitudinal study on the relationship between circulating PCSK9 and the prediction of clinical

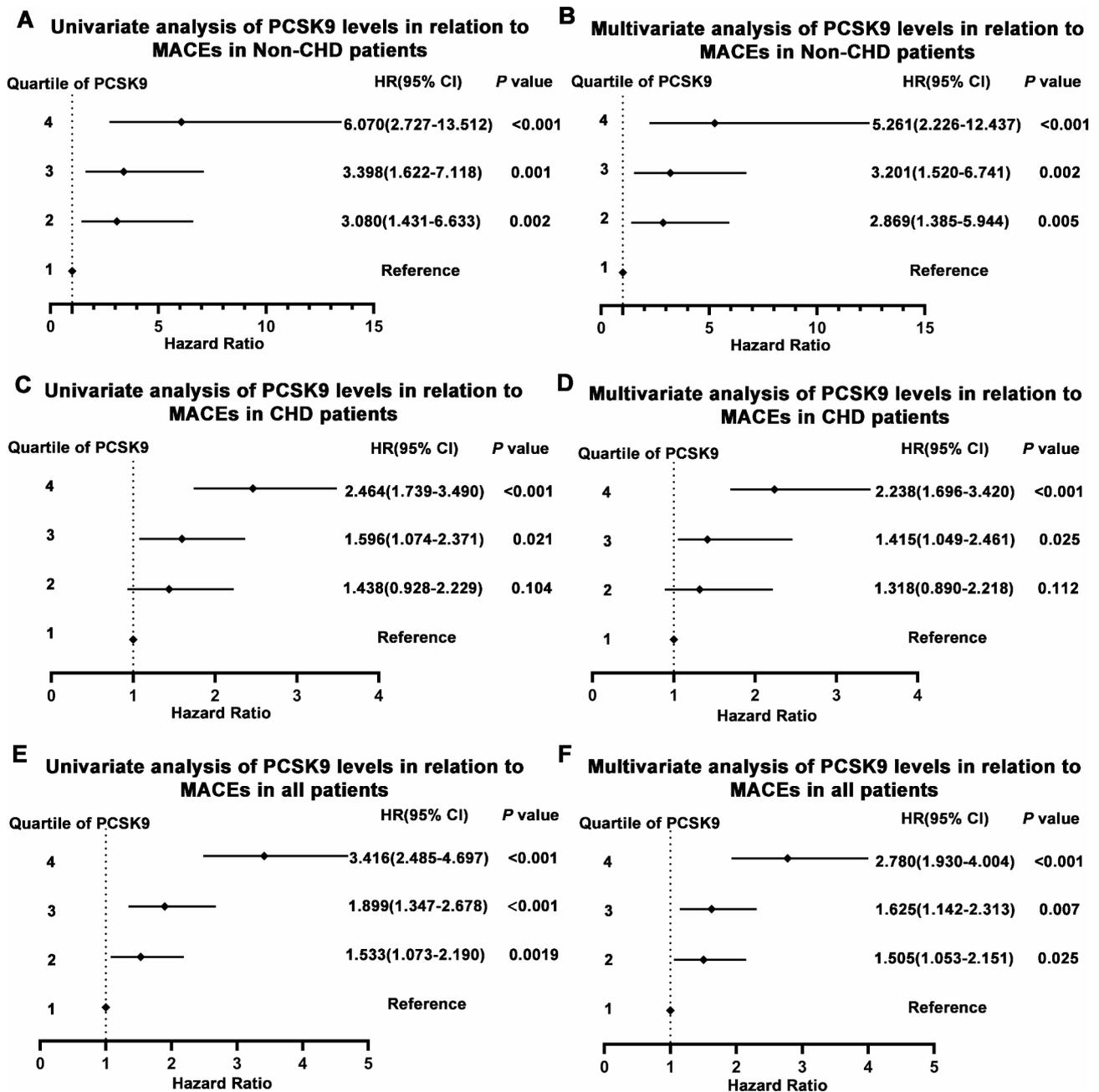


Fig. 5 Cox regression analysis of circulating PCSK9 quartile in different patients. (A) univariate Cox regression analysis in non-CHD group; (B) multivariate Cox regression analysis in non-CHD group; (C) univariate Cox regression analysis in CHD group; (D) multivariate Cox regression analysis in CHD group; (E) univariate Cox regression analysis in all participants; (F) multivariate Cox regression analysis in all participants

cardiovascular outcomes in Chinese individuals with T2DM. Previous studies reported increased plasma PCSK9 levels in patients with T2DM [18, 21, 22]. Moreover, among young participants, plasma PCSK9 levels are also increased in T1DM; as glycemic control deteriorates, plasma PCSK9 levels increase significantly [23]. Insulin resistance is one of the primary causes of diabetes. Previous studies have shown that insulin resistance and subsequent hyperinsulinemia can enhance plasma circulating

PCSK9 levels, and that the increase in plasma PCSK9 levels is related to poor glycemic control in patients with diabetes. Additionally, insulin resistance plays a crucial role in the homeostasis of PCSK9 in severe obesity. The PCSK9 controls the expression of LDLR on the surface of pancreatic cells, which may help maintain appropriate physiological balance to limit cholesterol overload in β cells. Pharmacological studies have shown that liraglutide can inhibit the expression of PCSK9 in HepG2 cells

and db/db mice via a hepatocyte nuclear factor 1 alpha-dependent mechanism. Notably, patients with diabetes accompanied by other metabolic disorders, especially atherosclerotic cardiovascular disease, have an increased probability of experiencing cardiovascular events [24, 25]. Additionally, atherosclerotic dyslipidemia plays a vital role in the prognosis of diabetes and atherosclerotic cardiovascular diseases [26]. Even with standard treatments, patients with CHD and diabetes have an increased cardiovascular risk. Therefore, further in-depth understanding of the cardiovascular risk in CHD patients with diabetes may help to prevent and better manage such patients [4, 8, 27].

PCSK9 binds to LDLR, leading to their intracellular degradation, thereby increasing plasma LDL-C levels and hyperlipidemia [11]. Soluble PCSK9 binds to the epidermal growth factor homologous domain of LDLR, which is mainly located on the surface of liver cells, thereby preventing it from adhering to LDL-C particles [28]. In addition, it promotes LDLR degradation by enhancing endocytosis and preventing recycling [29]. In addition to the classical pathway, PCSK9 regulates lipid metabolism via the intracellular endogenous PCSK9 regulation of ApoB expression and Lp(a) metabolism [30]. PCSK9 is associated with endothelial cell apoptosis, macrophage cholesterol efflux, intracellular mitochondrial dysfunction, and inflammation, all of which lead to metabolic disorders [27, 31]. Indeed, it has recently been suggested that high levels of circulating PCSK9 could directly promote the occurrence and progression of atherosclerosis, independently of the LDL-C levels [32]. Our data from the CHD group and non-CHD group, as well as the MACEs group and non-MACEs group, also support this viewpoint. In recent years, many studies have explored the role of PCSK9 in atherosclerosis; however, the predictive role of PCSK9 in cardiovascular outcomes remains controversial [33, 34]. PCSK9 expression is positively correlated with cardiovascular events in patients with acute coronary syndrome [35]. However, in a prospective cohort study on primary prevention, the results showed that baseline plasma level of PCSK9 could not predict future cardiovascular events [36]. Therefore, further evidence is required to support PCSK9 as a new predictor of MACEs, particularly in patients with CHD and T2DM.

PCSK9 potentially interacted with several proteins, including ApoB, ApoA1, APOE, LDLR, LRP1, SORT1 and so on. Interestingly, PCSK9 was related to SORT1, which was also named Sortilin, functions as a sorting receptor in the Golgi compartment and as a clearance receptor on the cell surface [37]. It could bind and mediate degradation of lipoprotein lipase that indicate that PCSK9 may be related to SORT1 in CHD severity, and MACEs in patients with T2DM [38, 39]. These results may guide future studies of the functional interactions

of PCSK9. In addition, GO term annotation showed that these proteins were mainly involved in cholesterol metabolic process, lipoprotein particle and lipoprotein particle receptor binding. A KEGG pathway analysis indicated enrichment in the cholesterol metabolism pathway. These could provide more information for us to further study the mechanism of PCSK9 in the severity of CHD and MACEs in patients with T2DM.

Previous studies have shown that PCSK9 is positively correlated with some traditional risk factors for atherosclerosis, and our study also confirmed a significant positive correlation between PCSK9 and TC and LDL-C. Previous studies have explored whether serum PCSK9 levels may be a new regulatory biomarker for the early diagnosis of atherosclerosis in T2DM. Furthermore, PCSK9 inhibitors can significantly reduce TC and LDL-C levels in patients with T2DM or atherosclerotic cardiovascular disease [40, 41]. Regardless of sex, the patients had high non-HDL-C and LDL-C levels. In the previous FOURIER and ODYSSEY trials, long-term follow-up showed that, the PCSK9 inhibitor alirocumab was more effective in reducing the occurrence of MACEs than the placebo control group [42, 43]. Currently, PCSK9 inhibitors are an effective treatment options for patients with type 2 diabetes and mixed dyslipidemia.

Several studies have shown that plasma PCSK9 levels are significantly higher in patients with CHD than in healthy individuals [20, 44, 45]. Existing epidemiological and clinical research results have shown that PCSK9 levels are high in patients with diabetes. In our study, in a group of Chinese Han T2DM patients, the plasma level of PCSK9 was positively correlated with the severity of coronary heart disease, as shown by the linear correlation results (diseased vessels: $r=0.35$, $p<0.001$; GS: $r=0.46$, $p<0.001$, significantly). To date, only Armentaro et al. have reported similar results in Caucasian populations [20]. It is important to emphasize how the genetic background affects clinical outcomes. The circulating plasma levels of PCSK9 vary significantly among individuals and may be influenced by genetic factors. Previous reports have shown differences in PCSK9 levels among people of different races, with Hispanics having higher levels than African-Americans and European-Americans [4]. In addition, Lakoski et al. studied the effect of PCSK9 allele sequence variation on plasma levels, and showed that African-Americans are heterozygous for PCSK9 non-sense mutations. Compared to people without invalid alleles, the median value of PCSK9 was low [44]. Through a six-year follow-up, our study not only proved that the PCSK9 level is proportional to the severity of CHD in Chinese Han T2DM patients, but also found that patients with a higher level of PCSK9 are more prone to MACEs.

Conclusion

Our study suggests a positive correlation between PCSK9 and multi-vessel CHD, and GS and indicates that, PCSK9 is an independent risk factor for CHD. Elevated PCSK9 levels are an independent predictor of MACEs in T2DM patients and T2DM patients with CHD.

This study has some limitations. First, only two centers participated in the study, which may have led to a selection bias in the results obtained. Some conclusions should be verified in more extensive multicenter studies. Second, our study lacked healthy individuals or those with pre-diabetes; therefore, we are unsure whether PCSK9 affects diabetes. Therefore, future studies should focus on this type of cohort. However, our data provide evidence for the clinical significance of PCSK9 levels in CHD and the risk of MACEs in T2DM patients.

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s13098-023-01238-z>.

Supplementary Material 1: Figure 1. Protein–protein interaction network of PCSK9 by using STRING

Supplementary Material 2: Figure 2. Enrichment analysis of PCSK9 functional networks in HNSC. Significantly enriched Gene Ontology annotations and Kyoto Encyclopedia of Genes and Genomes pathways of PCSK9. CC: Cellular component; BP: Biological process; MF: Molecular function

Supplementary Material 3: Baseline characteristics of type 2 diabetic patients with coronary heart disease at different circulating PCSK9 levels

Supplementary Material 4: The relationship between PCSK9 level and the MACEs outcomes in Non-CHD group

Supplementary Material 5: The relationship between PCSK9 level and the MACEs outcomes in CHD patients

Supplementary Material 6: The relationship between PCSK9 level and the MACEs outcomes in all patients

Supplementary Material 7: The relationship between PCSK9 level and the MACEs outcomes in male patients

Supplementary Material 8: The relationship between PCSK9 level and the MACEs outcomes in female patients

Supplementary Material 9: Multivariate analysis of PCSK9 levels in relation to MACEs in all patients

Acknowledgements

The authors thank all the staff and participants of this study for their important contributions.

Author contributions

J.H, J.X.G, A.M.Z, M.J, and M.S: the conception and design of the study. K.W, X.Z.Z, T.T.H, X.Q.Y, M.Y, Y.Y, S.S.L, N.Z, and J.J.H: acquisition of data, analysis and interpretation of data. J.H, J.X.G, K.W, A.M.Z, M.S, J.J.H and M.J: drafting the article and revising it critically for important intellectual content. J.H, X.Z.Z, J.J.H, M.S and M.J: final approval of the version to be submitted.

Funding

This study was supported by the Beijing Natural Science Foundation (7222194) and the Natural Science Foundation of Hunan Province (2021JJ41023).

Data availability

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Declarations

Ethics approval and consent to participate

This study complied with the Declaration of Helsinki and was approved by the Peking University People's Hospital Research Ethics Committee (No.2018PHB155-01). Written informed consent was obtained from all the enrolled patients.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

Author details

¹Department of traditional Chinese medicine, Peking University International Hospital, No. 1 Shengmingyuan Road, Zhongguancun Life Science Park, Changping District, Beijing 102206, P.R. China

²Department of Clinical Laboratory, Peking University People's Hospital, No. 11 Xizhimen South Street, Beijing 100044, P.R. China

³Department of Clinical Laboratory, Xiangya Hospital, Central South University, Changsha, P.R. China

⁴Department of Clinical Laboratory, Peking University International Hospital, Beijing, P.R. China

Received: 27 July 2023 / Accepted: 9 December 2023

Published online: 20 December 2023

References

- Verges B, Hassid J, Rouland A, Bouillet B, Simoneau I, Petit JM, et al. Liraglutide reduces plasma PCSK9 in patients with type 2 Diabetes not treated with statins. *Diabetes Metab.* 2022;48(2):101284.
- Pelletier RM, Layeghkhavidaki H, Seidah NG, Prat A, Vitale ML. PCSK9 contributes to the cholesterol, glucose, and Insulin2 homeostasis in Seminiferous Tubules and maintenance of Immunotolerance in Testis. *Front Cell Dev Biol.* 2022;10:889972.
- Furtado RHM, Fagundes AA Jr., Oyama K, Zelniker TA, Tang M, Kuder JF, et al. Effect of Evolocumab in patients with prior percutaneous coronary intervention. *Circ Cardiovasc Interv.* 2022;15(3):e011382.
- Ference BA, Robinson JG, Brook RD, Catapano AL, Chapman MJ, Neff DR, et al. Variation in PCSK9 and HMGR and risk of Cardiovascular Disease and Diabetes. *N Engl J Med.* 2016;375(22):2144–53.
- Gu JX, Huang J, Li SS, Zhou LH, Yang M, Li Y, et al. Elevated lipoprotein(a) and genetic polymorphisms in the LPA gene may predict cardiovascular events. *Sci Rep.* 2022;12(1):3588.
- Huang K, Liang Y, Wang K, Wu J, Luo H, Yi B. Influence of circulating nesfatin-1, GSH and SOD on insulin secretion in the development of T2DM. *Front Public Health.* 2022;10:882686.
- Wild S, Roglic G, Green A, Sicree R, King H. Global prevalence of Diabetes: estimates for the year 2000 and projections for 2030. *Diabetes Care.* 2004;27(5):1047–53.
- Lotta LA, Sharp SJ, Burgess S, Perry JRB, Stewart ID, Willems SM, et al. Association between Low-Density Lipoprotein Cholesterol-Lowering Genetic Variants and risk of type 2 Diabetes: a Meta-analysis. *JAMA.* 2016;316(13):1383–91.
- Brunham LR, Kruit JK, Verchere CB, Hayden MR. Cholesterol in islet dysfunction and type 2 Diabetes. *J Clin Invest.* 2008;118(2):403–8.
- Abifadel M, Varret M, Rabes JP, Allard D, Ouguerram K, Devillers M, et al. Mutations in PCSK9 cause autosomal dominant hypercholesterolemia. *Nat Genet.* 2003;34(2):154–6.
- Da Dalt L, Ruscica M, Bonacina F, Balzarotti G, Dhyani A, Di Cairano E, et al. PCSK9 deficiency reduces insulin secretion and promotes glucose intolerance: the role of the low-density lipoprotein receptor. *Eur Heart J.* 2019;40(4):357–68.

12. de Carvalho LSF, Campos AM, Sposito AC. Proprotein Convertase Subtilisin/Kexin type 9 (PCSK9) inhibitors and incident type 2 Diabetes: a systematic review and Meta-analysis with over 96,000 patient-years. *Diabetes Care*. 2018;41(2):364–7.
13. Ding Z, Wang X, Liu S, Shahanawaz J, Theus S, Fan Y, et al. PCSK9 expression in the ischaemic heart and its relationship to infarct size, cardiac function, and development of autophagy. *Cardiovasc Res*. 2018;114(13):1738–51.
14. Yang XQ, Mao YQ, Wang DX. More consideration of beta-cell function and PCSK9/LDLR axis. *Eur Heart J*. 2020;41(1):189.
15. Hu J, Zhang M, Gui L, Wan Q, Zhong J, Bai L, et al. PCSK9 suppresses M2-Like Tumor-Associated Macrophage polarization by regulating the secretion of OX40L from Hepatocellular Carcinoma Cells. *Immunol Invest*. 2022;51(6):1678–93.
16. Huang G, Lu X, Zhou H, Li R, Huang Q, Xiong X, et al. PCSK9 inhibition protects against myocardial ischemia-reperfusion injury via suppressing autophagy. *Microvasc Res*. 2022;142:104371.
17. Levstek T, Podkrajsek N, Rehberger Likozar A, Sebestjen M, Trebusak Podkrajsek K. The influence of treatment with PCSK9 inhibitors and variants in the CRP (rs1800947), TNFA (rs1800629), and IL6 (rs1800795) genes on the corresponding inflammatory markers in patients with very high lipoprotein(a) levels. *J Cardiovasc Dev Dis* 2022;9(5).
18. Momtazi AA, Banach M, Pirro M, Stein EA, Sahebkar A. PCSK9 and Diabetes: is there a link? *Drug Discov Today*. 2017;22(6):883–95.
19. Maliglowka M, Kosowski M, Hachula M, Cyrnek M, Buldak L, Basiak M et al. Insight into the Evolving Role of PCSK9. *Metabolites*. 2022;12(3).
20. Armentaro G, Carbone F, Cassano V, Liberale L, Minetti S, Bertolotto MB, et al. Serum proprotein convertase subtilisin/Kexin type 9 and vascular Disease in type 2 diabetic patients. *Eur J Clin Invest*. 2023;53(3):e13900.
21. Wang MM, Lu CF, Yan SQ, Wang BZ, Yesitayi G, Tian YL, et al. Association of genetic polymorphisms of PCSK9 with type 2 Diabetes in Uyur Chinese population. *BMC Cardiovasc Disord*. 2022;22(1):284.
22. Guo W, Gong Y, Li J, Qin P, Lu J, Li X, et al. Association of serum proprotein convertase subtilisin/kexin type 9 with early Atherosclerosis in newly diagnosed type 2 Diabetes Mellitus. *Nutr Metab Cardiovasc Dis*. 2019;29(8):815–21.
23. Bojanin D, Vekic J, Milenkovic T, Vukovic R, Zeljkovic A, Stefanovic A, et al. Association between proprotein convertase subtilisin/kexin 9 (PCSK9) and lipoprotein subclasses in children with type 1 Diabetes Mellitus: effects of glycemic control. *Atherosclerosis*. 2019;280:14–20.
24. Brunham LR, Kruit JK, Pape TD, Timmins JM, Reuwer AQ, Vasanji Z, et al. Beta-cell ABCA1 influences insulin secretion, glucose homeostasis and response to thiazolidinedione treatment. *Nat Med*. 2007;13(3):340–7.
25. Diabetes Prevention Program Research G. HbA1c as a predictor of Diabetes and as an outcome in the Diabetes prevention program: a randomized clinical trial. *Diabetes Care*. 2015;38(1):51–8.
26. Stone NJ, Robinson JG, Lichtenstein AH, Bairey Merz CN, Blum CB, Eckel RH, et al. 2013 ACC/AHA guideline on the treatment of blood cholesterol to reduce atherosclerotic cardiovascular risk in adults: a report of the American College of Cardiology/American Heart Association Task Force on Practice guidelines. *Circulation*. 2014;129(25 Suppl 2):1–45.
27. Wolf A, Kutsche HS, Schreckenber R, Weber M, Li L, Rohrbach S, et al. Auto-crine effects of PCSK9 on cardiomyocytes. *Basic Res Cardiol*. 2020;115(6):65.
28. Dong B, Li H, Singh AB, Cao A, Liu J. Inhibition of PCSK9 transcription by berberine involves down-regulation of hepatic HNF1alpha protein expression through the ubiquitin-proteasome degradation pathway. *J Biol Chem*. 2015;290(7):4047–58.
29. Path G, Perakakis N, Mantzoros CS, Seufert J. PCSK9 inhibition and cholesterol homeostasis in insulin producing beta-cells. *Lipids Health Dis*. 2022;21(1):138.
30. Poirier S, Mayer G, Benjannet S, Bergeron E, Marcinkiewicz J, Nassoury N, et al. The proprotein convertase PCSK9 induces the degradation of low density lipoprotein receptor (LDLR) and its closest family members VLDLR and ApoER2. *J Biol Chem*. 2008;283(4):2363–72.
31. Marmugi A, Parnis J, Chen X, Carmichael L, Hardy J, Mannan N, et al. Sorcin links pancreatic beta-cell lipotoxicity to ER Ca2+ stores. *Diabetes*. 2016;65(4):1009–21.
32. Toscano A, Cinquegrani M, Scuruchi M, Di Pino A, Piro S, Ferrara V et al. PCSK9 plasma levels are Associated with mechanical vascular impairment in familial hypercholesterolemia subjects without a history of atherosclerotic Cardiovascular Disease: results of six-Month Add-On PCSK9 inhibitor therapy. *Biomolecules* 2022;12(4).
33. El Khoury P, Roussel R, Fumeron F, Abou-Khalil Y, Velho G, Mohammadi K, et al. Plasma proprotein-convertase-subtilisin/kexin type 9 (PCSK9) and cardiovascular events in type 2 Diabetes. *Diabetes Obes Metab*. 2018;20(4):943–53.
34. Vlachopoulos C, Terentes-Printzios D, Georgiopoulos G, Skoumas I, Koutagiari I, Ioakeimidis N, et al. Prediction of cardiovascular events with levels of proprotein convertase subtilisin/kexin type 9: a systematic review and meta-analysis. *Atherosclerosis*. 2016;252:50–60.
35. Gencer B, Montecucco F, Nanchen D, Carbone F, Klingenberg R, Vuilleumier N, et al. Prognostic value of PCSK9 levels in patients with acute coronary syndromes. *Eur Heart J*. 2016;37(6):546–53.
36. Ridker PM, Rifai N, Bradwin G, Rose L. Plasma proprotein convertase subtilisin/kexin type 9 levels and the risk of first cardiovascular events. *Eur Heart J*. 2016;37(6):554–60.
37. Ni X, Morales CR. The lysosomal trafficking of acid sphingomyelinase is mediated by sortilin and mannose 6-phosphate receptor. *Traffic*. 2006;7(7):889–902.
38. Nielsen MS, Jacobsen C, Olivecrona G, Gliemann J, Petersen CM. Sortilin/neurotensin receptor-3 binds and mediates degradation of lipoprotein lipase. *J Biol Chem*. 1999;274(13):8832–6.
39. Gustafsen C, Kjolby M, Nyegaard M, Mattheisen M, Lundhede J, Buttenschon H, et al. The hypercholesterolemia-risk gene SORT1 facilitates PCSK9 secretion. *Cell Metab*. 2014;19(2):310–8.
40. Raber L, Ueki Y, Otsuka T, Losdat S, Haner JD, Lonborg J, et al. Effect of Alirocumab added to high-intensity statin therapy on coronary Atherosclerosis in patients with Acute Myocardial Infarction: the PACMAN-AMI Randomized Clinical Trial. *JAMA*. 2022;327(18):1771–81.
41. Ray KK, Colhoun HM, Szarek M, Baccara-Dinet M, Bhatt DL, Bittner VA, et al. Effects of alirocumab on cardiovascular and metabolic outcomes after acute coronary syndrome in patients with or without Diabetes: a prespecified analysis of the ODYSSEY OUTCOMES randomised controlled trial. *Lancet Diabetes Endocrinol*. 2019;7(8):618–28.
42. Schwartz GG, Gabriel Steg P, Bhatt DL, Bittner VA, Diaz R, Goodman SG, et al. Clinical efficacy and safety of Alirocumab after Acute Coronary Syndrome according to Achieved Level of low-density lipoprotein cholesterol: a propensity score-matched analysis of the ODYSSEY OUTCOMES trial. *Circulation*. 2021;143(11):1109–22.
43. Colhoun HM, Ginsberg HN, Robinson JG, Leiter LA, Muller-Wieland D, Henry RR, et al. No effect of PCSK9 inhibitor alirocumab on the incidence of Diabetes in a pooled analysis from 10 ODYSSEY phase 3 studies. *Eur Heart J*. 2016;37(39):2981–9.
44. Lakoski SG, Lagace TA, Cohen JC, Horton JD, Hobbs HH. Genetic and metabolic determinants of plasma PCSK9 levels. *J Clin Endocrinol Metab*. 2009;94(7):2537–43.
45. Kouvari M, Chrysohoou C, Skoumas J, Pitsavos C, Panagiotakos DB, Mantzoros CS, et al. The presence of NAFLD influences the transition of metabolically healthy to metabolically unhealthy obesity and the ten-year Cardiovascular Disease risk: a population-based cohort study. *Metabolism*. 2022;128:154893.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.